

Exploration and Integration of Heterogeneous Biological Data Sets with mixOmics



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Outline

- **Introduction:** interdisciplinarity, data integration, answer a question
- **Tools:** mixOmics R package, workflow
- **Methods:** understand PCA, extend to integration problems, sparsity, vertical integration
- **Examples:** simulated toy examples, liver toxicity data set

Interdisciplinarity

*The biological sciences are **today** in the process of changing from being primarily descriptive **to being very much quantitative**. As a result, biologists find themselves **confronted more and more with large amounts of numerical data** [...]. But the mere collecting and recording of data achieve nothing; having been collected, they must be **investigated to see what information may be contained concerning the biological problem** at hand.[...]*

*Frequently, however, biologists have to subject their data to more complex calculations, requiring procedures that **involve mathematical details beyond their general experience**. In order to carry out the mathematics the biologist in this situation must either **learn the procedures himself**, or at least **learn something of the language of mathematics**, that he may **communicate satisfactorily with the mathematician** whose aid he enlists.*

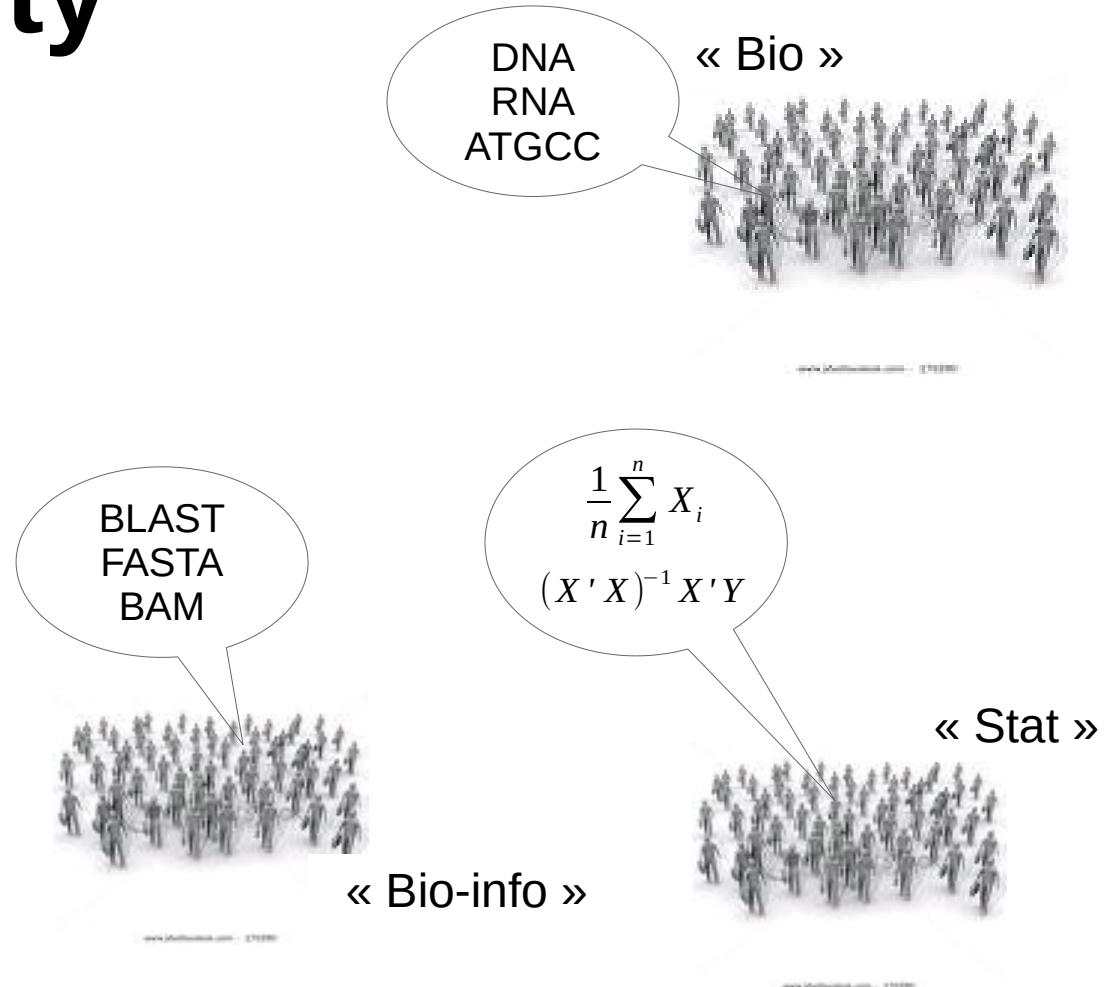
S.R Searle ([1966](#))

Matrix Algebra for the biological sciences

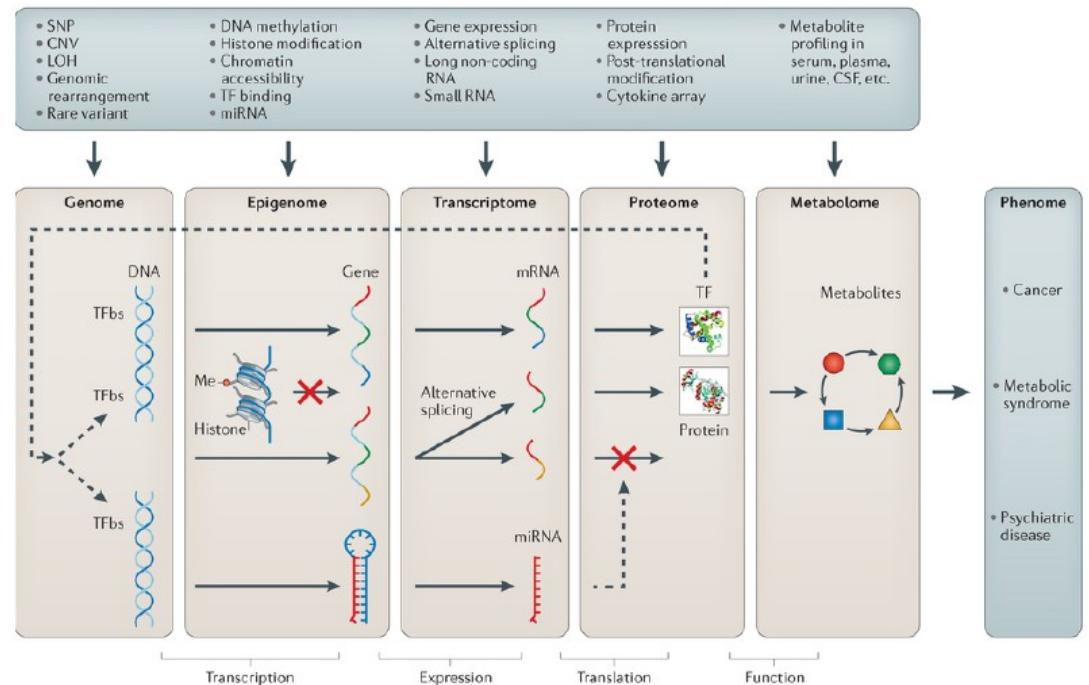
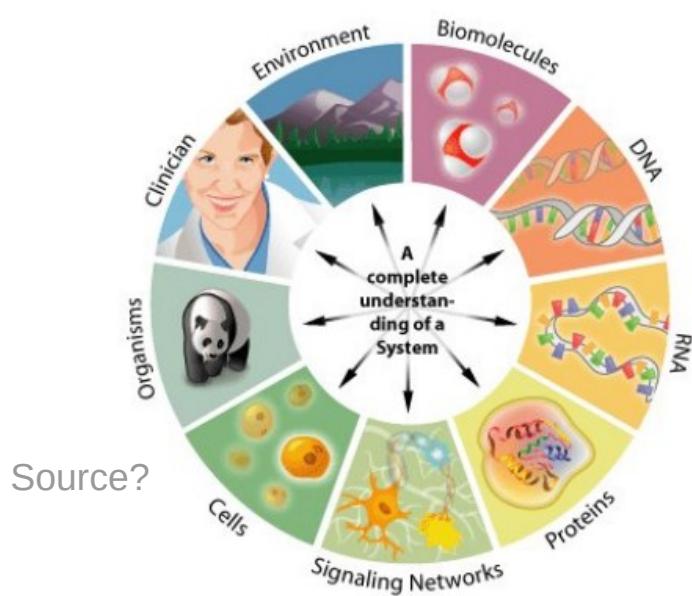
Interdisciplinarity

- Nearly unlimited quantity of data from multiple and heterogeneous sources
- Computational issues to foresee
- Biological interpretation for validation
- Keep pace with new technologies

A close interaction between statisticians, bioinformaticians and molecular biologists is essential to provide meaningful results



Data integration



From Ritchie et al. (2015), *Nature reviews. Genetics*
Methods of integrating data to uncover genotype-phenotype interactions.

Generally, data integration can be defined as the process of combining data residing in diverse sources to provide users with a comprehensive view of such data. There is no universal approach to data integration, and many techniques are still evolving.

From Schneider, M. V., & Jimenez, R. C. (2012). Teaching the Fundamentals of Biological Data Integration Using Classroom Games. PLoS Computational Biology, 8(12)

Data integration with statistics

Goal: extract knowledge from data



Answer a question

THE FUTURE OF DATA ANALYSIS¹

By JOHN W. TUKEY

Princeton University and Bell Telephone Laboratories

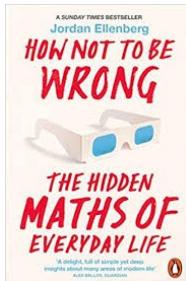
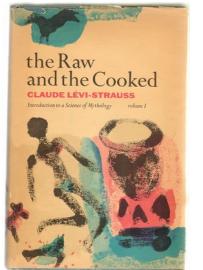
Received July 1, 1961.

¹ Prepared in part in connection with research sponsored by the Army Research Office through Contract DA36-034-ORD-2297 with Princeton University. Reproduction in whole or part is permitted for any purpose of the United States Government.

*Far better an approximate answer to **the right question** [...], than an exact answer to the wrong question [...].*

*The scientific mind does not so much provide the right answers as **ask the right questions**.*

C. Lévi-Strauss. *The Raw and the Cooked* (1964)



[...] in order to give a sensible answer, you need to know more than just numbers [...] It's **only after you've started to formulate these questions** that you take out the calculator. But **at that point the real mental work is already finished**. Dividing one number by another is mere computation; figuring out what you should divide by what is mathematics.

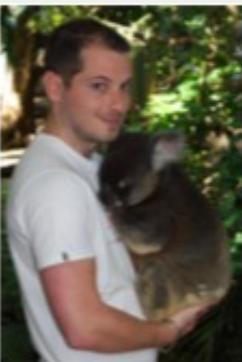
Tool(s)

- Package for R 
r-project.org
- Freely available on
Bioconductor 
bioconductor.org/packages/release/bioc/html/mixOmics.html
- Web site mixomics.org
- Forum
mixomics-users.discourse.group



mixOmics facebook

- Core team



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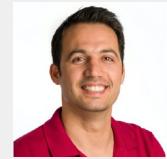


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- Many users and trainees



mixOmics workflow

1) Run a method: `pca(DataX)`, `sPCA(DataX, keepX=c(...))`, `pls(DataX, DataY)`,
`sPLS(DataX, DataY, keepX=c(...), keepY=c(...))`, `PLSDA(DataX, Categ)`, `sPLSDA(DataX, Categ, keepX=c(...))`, `block.pls(list(...))`, `block.sPLS(list(...))`, `block.PLSDA(list(...), Categ)`, `block.sPLSDA(list(...), Categ, keepX=list(...))`, `mint.pca(DataX, study=...)`, `mint.PLSDA(DataX, Categ, study=...)`, ...

Optional argument for compositional data (microbial dataset): `logratio = 'CLR'`

2) Represent individuals: `plotIndiv()`

3) Represent variables: `plotVar()`, `plotLoadings()`, `cim()`, `network()`

Overview of statistical methods

- **Multivariate unsupervised**

One numerical dataset `pca()`, `spca()`



- **Multivariate supervised**

One numerical dataset and one categorical variable `plsda()`, `splsda()`



Numerical

Categorical

- **Multi-block unsupervised**

Several numerical datasets, same samples

`pls()`, `spls()`, `block.pls()`, `block.spls()`



- **Multi-block supervised**

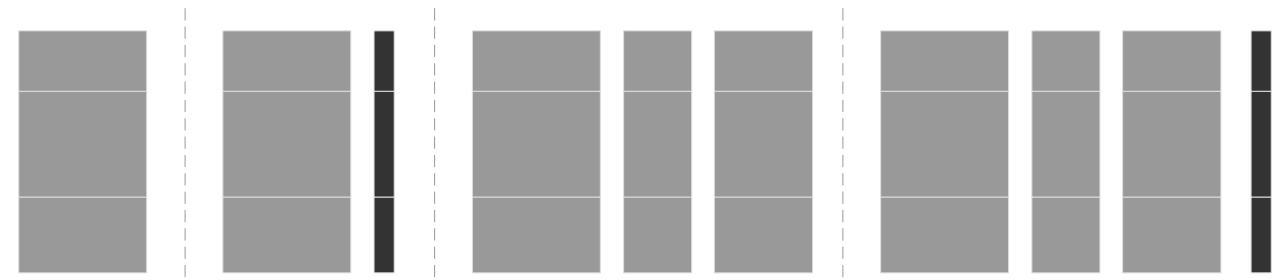
Several numerical datasets and one categorical variable, same samples
`block.plsda()`, `block.splsda()`



- **Multi-group analyses**

Same as above with samples divided pre-defined in groups (batch, study...)

`mint.pca()`, `mint.plsda()`, `mint.splsda()`,
`mint.block.pls()`, `mint.block.spls()`,
`mint.block.plsda()`, `mint.block.splda()`



Methods

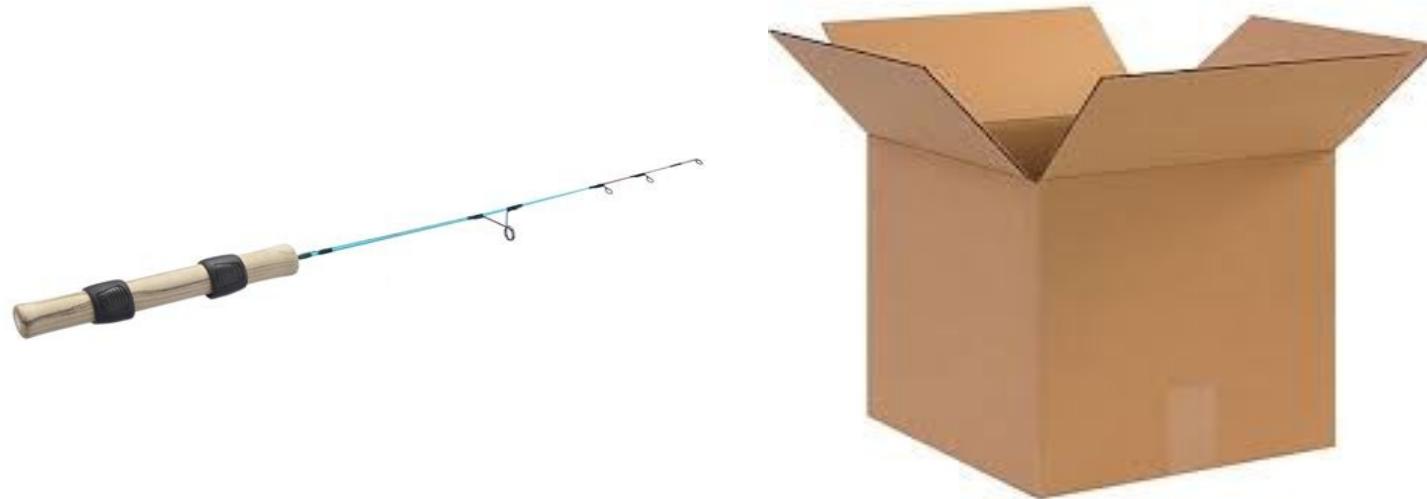
- Understand Principal Component Analysis
- Extend to integration problems
- Sparsity



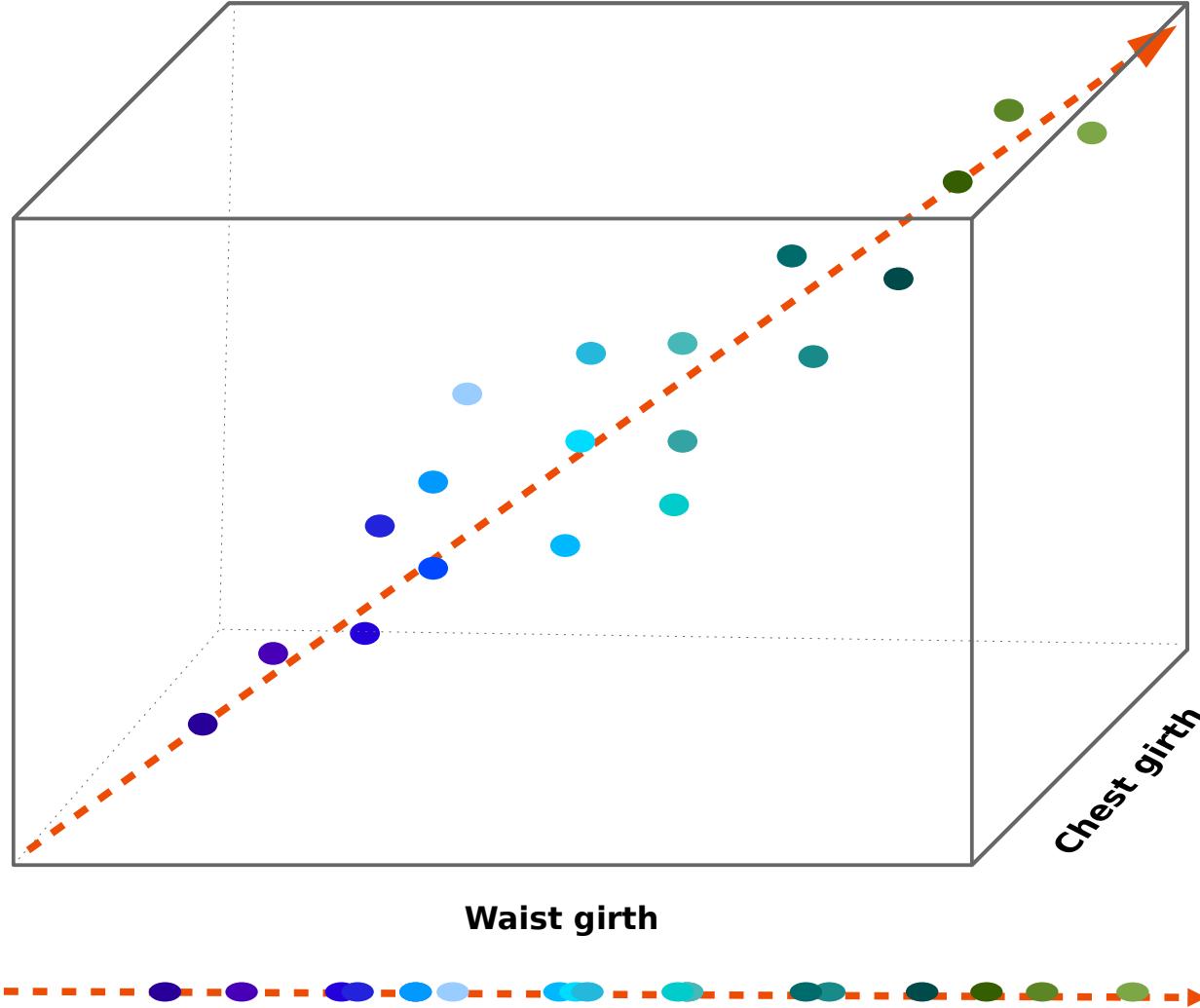
The Batman principle: *It's not who I am underneath, but what I do that defines me.*
From Batman Begins
www.youtube.com/watch?v=PmwLPU5H6_Q

Understand PCA

Teasing: Would you use a cubic box
to pack a fishing rod?



PCA



Do we need 3 dimensions to represent 'standard' individuals?

=

Do we need a cubic box to pack a fishing rod?

1st Principal Component:
«beefyness»

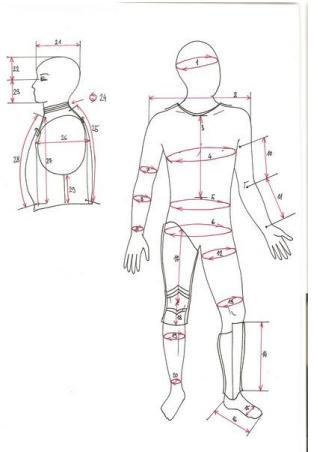
PCA: (verbose) comments

- The measurements are rather **strongly correlated**. Indeed, one can assume that a person with a high shoulder girth will also have high chest girth. In these conditions, the information brought by the 3 variables are **redundant**. Graphically, in the cube determined by shoulder girth, chest girth and waist girth, there are nearly empty areas. One variable calculated as a **combination** of these 3 variables (represented as the dotted arrow) would be enough to represent the individuals with a **minimal loss in information** because all the points are located along these direction that is the first principal component.
- PCA allows to determine the sub-spaces of lower dimension than the initial space on which the projection of the individuals is the **least modified**, that is to say, the sub-spaces that retain the **greatest part of the information** (i.e. **variability**).
- The principle of PCA consists in finding a direction (the first PC), calculated as a **linear combination of the initial variables**, such that the **variance** of the points around this direction is **maximal**. Iterate this process in orthogonal directions to determine the following principal components. The number of PC that can be calculated is equal to the number of initial variables.
- Concerning the variables, the PCA keeps at best the **correlation structure** between the initial variables.

A toy example

- 20 individuals
- 5 variables

s.g : shoulder girth (cm)
 c.g : chest girth (cm)
 w.g : waist girth (cm)
 w : weight (kg)
 h : height (cm)



Id	s.g	c.g	w.g	w	h
I1	106.2	89.5	71.5	65.6	174.0
I2	110.5	97.0	79.0	71.8	175.3
I3	115.1	97.5	83.2	80.7	193.5
I4	104.5	97.0	77.8	72.6	186.5
I5	107.5	97.5	80.0	78.8	187.2
I6	119.8	99.9	82.5	74.8	181.5
I7	123.5	106.9	82.0	86.4	184.0
I8	120.4	102.5	76.8	78.4	184.5
I9	111.0	91.0	68.5	62.0	175.0
I10	119.5	93.5	77.5	81.6	184.0
I11	105.0	89.0	71.2	67.3	169.5
I12	100.2	94.1	79.6	75.5	160.0
I13	99.1	90.8	77.9	68.2	172.7
I14	107.6	97.0	69.6	61.4	162.6
I15	104.0	95.4	86.0	76.8	157.5
I16	108.4	91.8	69.9	71.8	176.5
I17	99.3	87.3	63.5	55.5	164.4
I18	91.9	78.1	57.9	48.6	160.7
I19	107.1	90.9	72.2	66.4	174.0
I20	100.5	97.1	80.4	67.3	163.8

First computations

Raw data

Id	s.g	c.g	w.g	w	h
I1	106.2	89.5	71.5	65.6	174.0
I2	110.5	97.0	79.0	71.8	175.3
I3	115.1	97.5	83.2	80.7	193.5
I4	104.5	97.0	77.8	72.6	186.5
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I19	107.1	90.9	72.2	66.4	174.0
I20	100.5	97.1	80.4	67.3	163.8

Bivariate analysis

Covariance matrix

	s.g	c.g	w.g	w	h
s.g	68.6	37.7	28.1	55.3	61.2
c.g	37.7	37.5	33.9	45.7	32.4
w.g	28.1	33.9	50.8	56.6	27.7
w	55.3	45.7	56.6	85.7	59.5
h	61.2	32.4	27.7	59.5	109.3

Pearson correlation matrix

	s.g	c.g	w.g	w	h
s.g	1.0	0.7	0.5	0.7	0.7
c.g	0.7	1.0	0.8	0.8	0.5
w.g	0.5	0.8	1.0	0.9	0.4
w	0.7	0.8	0.9	1.0	0.6
h	0.7	0.5	0.4	0.6	1.0

Univariate analysis

Mean	108.1	94.2	75.3	70.6	174.4
Variance	68.6	37.5	50.8	85.7	109.3

$$68.6 + 37.5 + 50.8 + 85.7 + 109.3 = 351.9$$

351.9 represents the quantity of information contained in the data.

The core of PCA

Coefficients of linear combination (or loadings)

	PC1	PC2	PC3	PC4	PC5
shoulder.g	0.45	-0.16	0.78	-0.18	0.36
chest.g	0.32	0.25	0.26	0.72	-0.49
waist.g	0.34	0.53	-0.33	0.24	0.66
weight	0.54	0.36	-0.17	-0.60	-0.44
height	0.54	-0.70	-0.43	0.17	0.02

PC1 = 0.45*shoulder.g + 0.32*chest.g + 0.34*waist.g + 0.54*weight + 0.54*height

PC2 = -0.16*shoulder.g + 0.25*chest.g + 0.53*waist.g + 0.36*weight - 0.70*height

...



What is underneath? Bruce Wayne Eigen decomposition of the covariance matrix.

Around the core

Centered data

Id	s.g	c.g	w.g	w	h
I1	-1.9	-4.7	-3.8	-5.0	-0.4
I2	2.4	2.8	3.7	1.2	0.9
I3	7.0	3.3	7.9	10.1	19.1
I4	-3.6	2.8	2.5	2.0	12.1
I5	-0.6	3.3	4.7	8.2	12.8
I6	11.7	5.7	7.2	4.2	7.1
I7	15.4	12.7	6.7	15.8	9.6
I8	12.3	8.3	1.5	7.8	10.1
I9	2.9	-3.2	-6.8	-8.6	0.6
I10	11.4	-0.7	2.2	11.0	9.6
I11	-3.1	-5.2	-4.1	-3.3	-4.9
I12	-7.9	-0.1	4.2	4.9	-14.4
I13	-9.0	-3.4	2.6	-2.4	-1.7
I14	-0.5	2.8	-5.8	-9.2	-11.8
I15	-4.1	1.2	10.7	6.2	-16.9
I16	0.3	-2.4	-5.4	1.2	2.1
I17	-8.8	-6.9	-11.8	-15.1	-10.0
I18	-16.2	-16.1	-17.4	-22.0	-13.7
I19	-1.0	-3.3	-3.1	-4.2	-0.4
I20	-7.6	2.9	5.1	-3.3	-10.6

$$\text{Ex: } -6.50 = 0.45*(-1.9) + 0.32*(-4.7) + 0.34*(-3.8) + 0.54*(-5) + 0.54*(-0.4)$$

	PC1	PC2	PC3	PC4	PC5
I1	-6.50	-4.48	-0.37	-1.03	1.27
I2	4.40	2.04	0.81	1.87	1.38
I3	22.66	-5.94	-6.18	0.11	1.97
I4	7.78	-5.24	-8.38	4.10	-1.74
I5	13.73	-2.67	-8.02	0.82	-2.15
I6	15.67	-0.15	4.49	2.33	4.40
I7	26.99	3.19	6.29	0.04	-3.08
I8	18.41	-3.43	5.63	1.09	-1.96
I9	-6.25	-8.48	4.97	0.79	1.86
I10	16.78	-3.67	1.99	-7.08	1.22
I11	-8.83	-0.78	0.28	-3.02	0.07
I12	-7.28	15.41	-2.31	-3.00	-2.35
I13	-6.45	2.25	-7.60	0.95	1.15
I14	-12.51	2.68	8.91	4.27	-1.53
I15	-3.65	20.76	-0.30	-2.45	1.99
I16	-0.63	-4.62	0.34	-3.46	-2.80
I17	-23.61	-5.07	2.20	1.19	-1.15
I18	-37.50	-9.07	-1.33	-1.89	-0.02
I19	-4.98	-3.61	0.33	-0.50	1.02
I20	-8.24	10.89	-1.74	4.86	0.44

Apply loadings



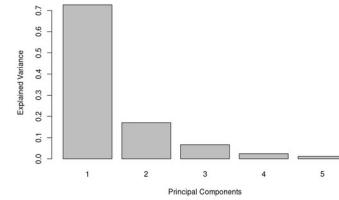
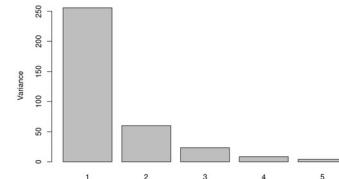
255.7 is the greatest variance we can obtain with a linear combination of the initial variables.

Mean	0	0	0	0	0	
Var.	255.7	60.2	23.5	8.6	4.0	= 351.9

Graphical outputs (1/3)

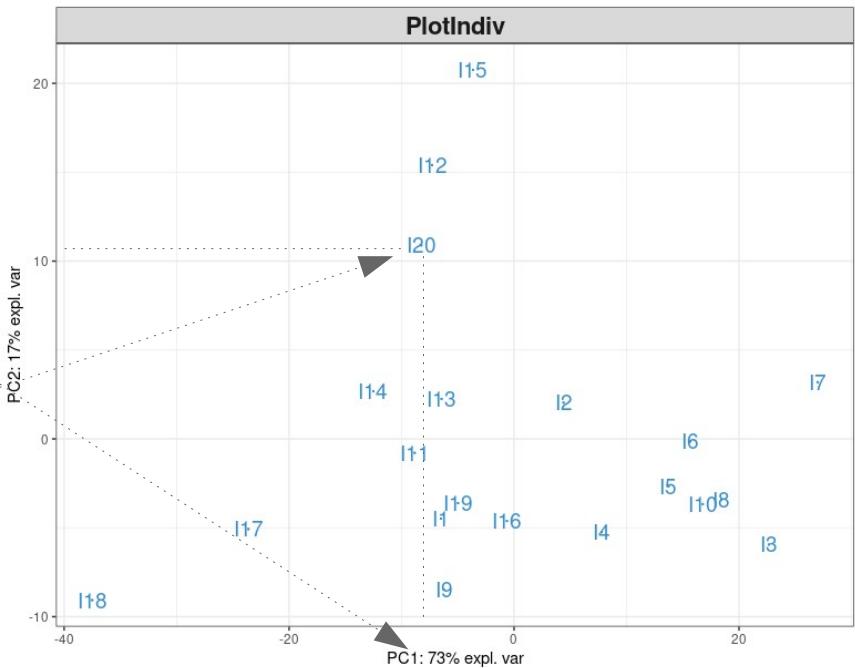
	PC1	PC2	PC3	PC4	PC5
Variance	255.7	60.2	23.5	8.6	4.0
% variance	72.6	17.1	6.7	2.4	1.1

Screeplot



	PC1	PC2	PC3	PC4	PC5
I1	-6.50	-4.48	-0.37	-1.03	1.27
I2	4.40	2.04	0.81	1.87	1.38
I3	22.66	-5.94	-6.18	0.11	1.97
I4	7.78	-5.24	-8.38	4.10	-1.74
I5	13.73	-2.67	-8.02	0.82	-2.15
I6	15.67	-0.15	4.49	2.33	4.40
I7	26.99	3.19	6.29	0.04	-3.08
I8	18.41	-3.43	5.63	1.09	-1.96
I9	-6.25	-8.48	4.97	0.79	1.86
I10	16.78	-3.67	1.99	-7.08	1.22
I11	-8.83	-0.78	0.28	-3.02	0.07
I12	-7.28	15.41	-2.31	-3.00	-2.35
I13	-6.45	2.25	-7.60	0.95	1.15
I14	-12.51	2.68	8.91	4.27	-1.53
I15	-3.65	20.76	-0.30	-2.45	1.99
I16	-0.63	-4.62	0.34	-3.46	-2.80
I17	-23.61	-5.07	2.20	1.19	-1.15
I18	-37.50	-9.07	-1.33	-1.89	-0.02
I19	-4.98	-3.61	0.33	-0.50	1.02
I20	-8.24	10.89	-1.74	4.86	0.44

Individual plot



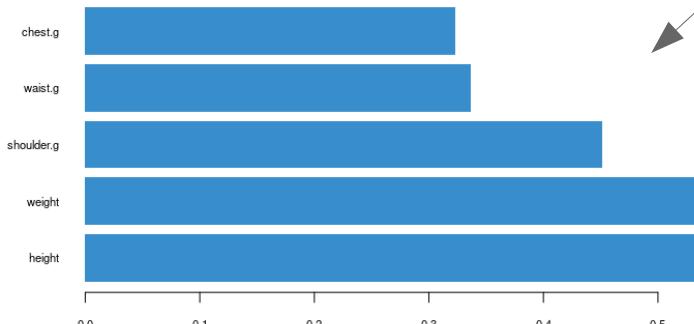
Graphical outputs (2/3)

Loadings

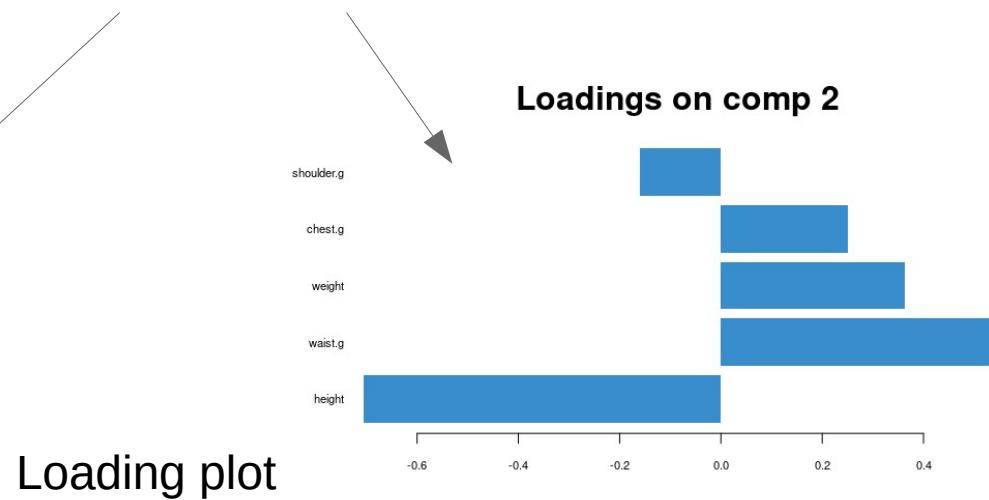
shoulder.g
chest.g
waist.g
weight
height

PC1	PC2
0.45	-0.16
0.32	0.25
0.34	0.53
0.54	0.36
0.54	-0.70

Loadings on comp 1



Loadings on comp 2



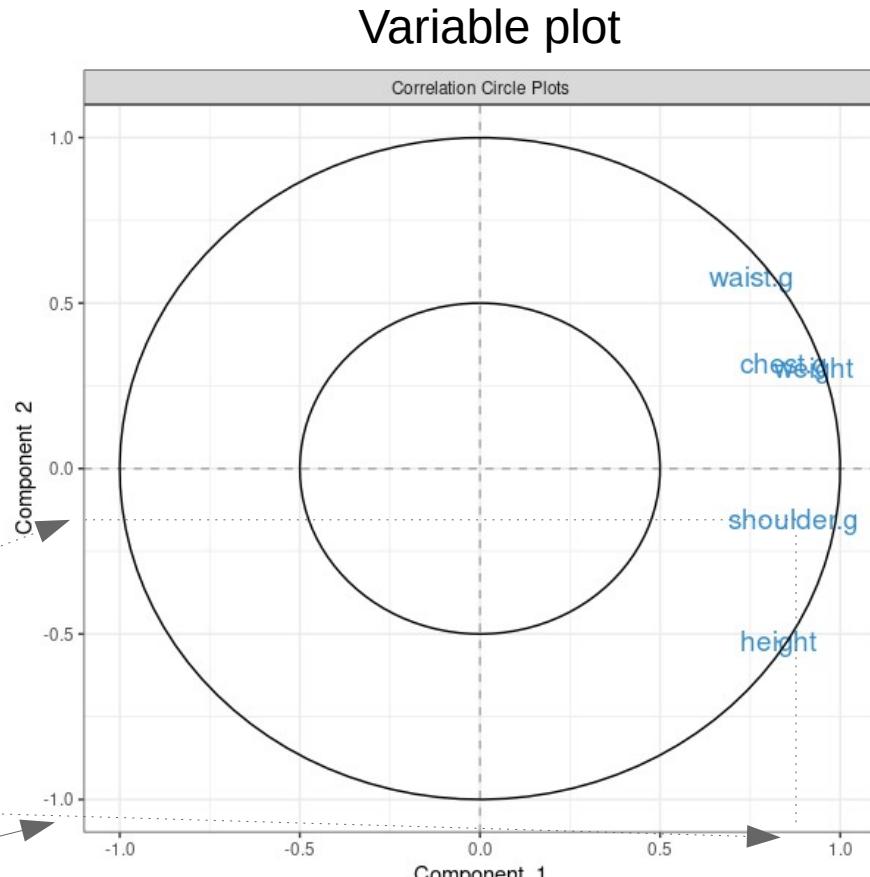
Graphical outputs (3/3)

Id	s.g	c.g	w.g	w	h		PC1	PC2
I1	106.2	89.5	71.5	65.6	174.0	I1	-6.50	-4.48
I2	110.5	97.0	79.0	71.8	175.3	I2	4.40	2.04
I3	115.1	97.5	83.2	80.7	193.5	I3	22.66	-5.94
I4	104.5	97.0	77.8	72.6	186.5	I4	7.78	-5.24
I5	107.5	97.5	80.0	78.8	187.2	I5	13.73	-2.67
I6	119.8	99.9	82.5	74.8	181.5	I6	15.67	-0.15
I7	123.5	106.9	82.0	86.4	184.0	I7	26.99	3.19
I8	120.4	102.5	76.8	78.4	184.5	I8	18.41	-3.43
I9	111.0	91.0	68.5	62.0	175.0	I9	-6.25	-8.48
I10	119.5	93.5	77.5	81.6	184.0	I10	16.78	-3.67
I11	105.0	89.0	71.2	67.3	169.5	I11	-8.83	-0.78
I12	100.2	94.1	79.6	75.5	160.0	I12	-7.28	15.41
I13	99.1	90.8	77.9	68.2	172.7	I13	-6.45	2.25
I14	107.6	97.0	69.6	61.4	162.6	I14	-12.51	2.68
I15	104.0	95.4	86.0	76.8	157.5	I15	-3.65	20.76
I16	108.4	91.8	69.9	71.8	176.5	I16	-0.63	-4.62
I17	99.3	87.3	63.5	55.5	164.4	I17	-23.61	-5.07
I18	91.9	78.1	57.9	48.6	160.7	I18	-37.50	-9.07
I19	107.1	90.9	72.2	66.4	174.0	I19	-4.98	-3.61
I20	100.5	97.1	80.4	67.3	163.8	I20	-8.24	10.89

$\text{cor}(s.g, \text{PC1}) = 0.87$
 $\text{cor}(s.g, \text{PC2}) = -0.15$

$\text{cor}(c.g, \text{PC1}) = 0.84$
 $\text{cor}(c.g, \text{PC2}) = 0.32$
 ...

	PC1	PC2
shoulder.g	0.87	-0.15
chest.g	0.84	0.32
waist.g	0.75	0.58
weight	0.92	0.30
height	0.83	-0.52

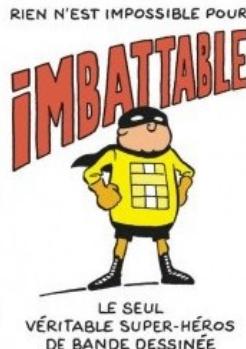


Focus on individual plot

- To interpret the graphical results of PCA must be done keeping in mind that one is looking at a projection on a plane (or in a volume for 3D representation)
- Be careful when interpreting visual proximities
- Illustration in comics with the only true super-heros ...

Scenario & illustration: Pascal Jousselin
Colour: Laurence Croix

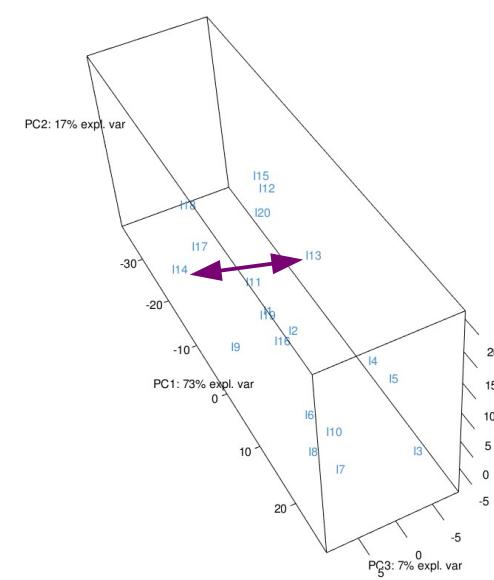
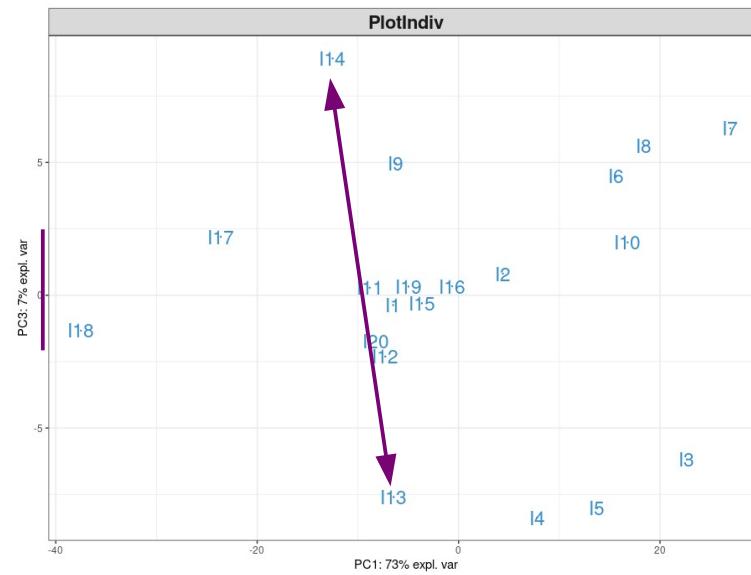
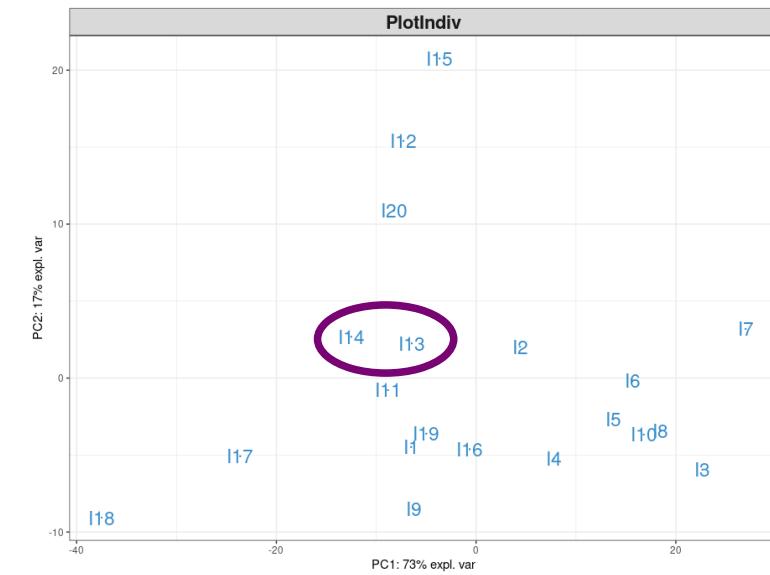
pjousselin.free.fr



Focus on individual plot



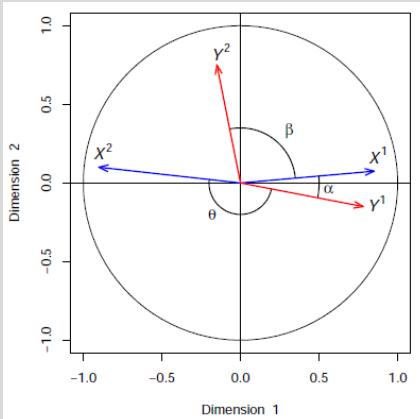
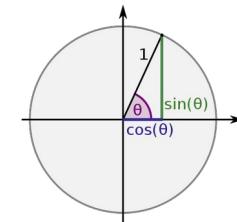
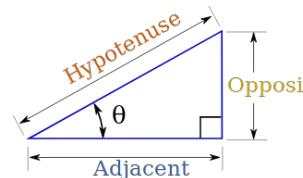
I13	99.1	90.8	77.9	68.2	172.7
I14	107.6	97.0	69.6	61.4	162.6



Focus on variable plot

Correlation \leftrightarrow cosine

Remember trigonometry and right triangles:



The correlation between two variables is represented as:

- An acute angle ($\cos(\alpha) > 0$) if it is positive
- An obtuse angle ($\cos(\theta) < 0$) if it is negative
- A right angle ($\cos(\beta) \approx 0$) if it is near zero

Graphical outputs: summary

Screeplot

- How many components?
- 90% with 2 Pcs, 97% with 3PCs, 100% with 5PCs

Individual plot

- ‘Natural’ clusters, outliers...
- Caution: visual proximities

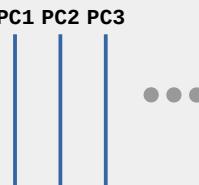
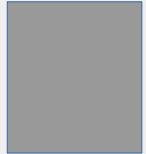


Variable plot, loading plot

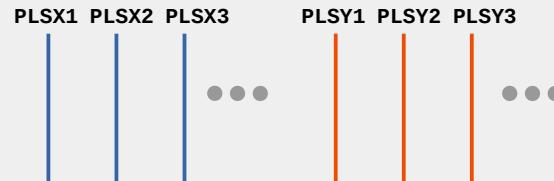
- Correlation between variables
- Interpret components: PC1 « beefyness », PC2 « fatness, rotundity »

Extension to integration problems

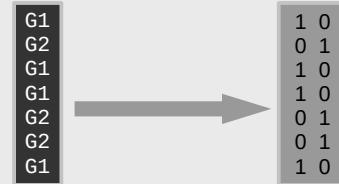
PCA
 $\max \text{var}(\text{PC}_i)$



PLS, PLS-DA
 $\max \text{cov}(\text{PLS}_x, \text{PLS}_y)$



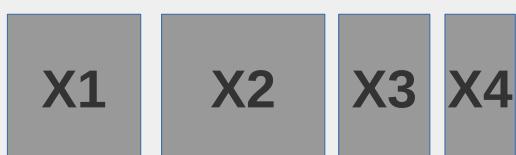
The trick for discriminant analyses:
 convert a factor into a numeric
 (dummy) matrix



PLS-DA → PLS

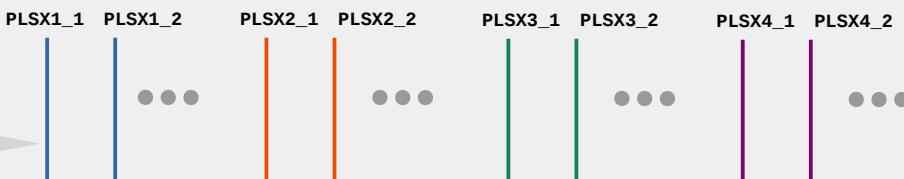
Generalized PLS, PLS-DA

$\max \{ c_{12} \cdot \text{cov}(\text{PLS}_{x_1}, \text{PLS}_{x_2}) +$
 $c_{13} \cdot \text{cov}(\text{PLS}_{x_1}, \text{PLS}_{x_3}) +$
 $c_{14} \cdot \text{cov}(\text{PLS}_{x_1}, \text{PLS}_{x_4}) +$
 $c_{23} \cdot \text{cov}(\text{PLS}_{x_2}, \text{PLS}_{x_3}) +$
 $\dots \}$



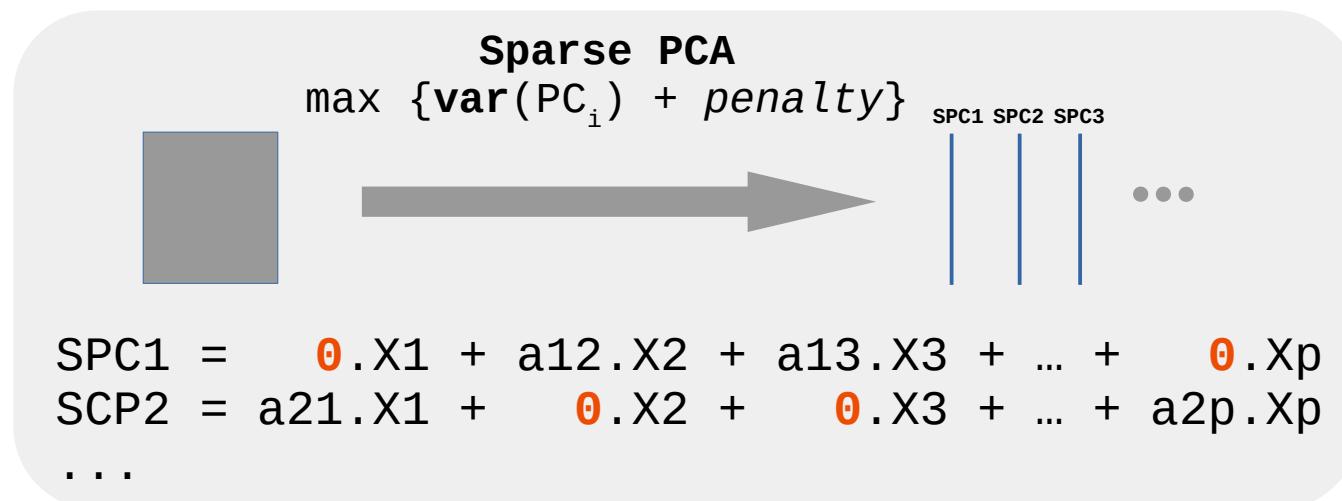
$\dots \}$

c_{ij} can be set by the user through a design matrix



Sparsity

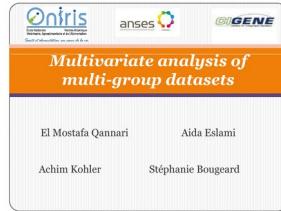
- High throughput experiments: too many variables, noisy or irrelevant depending on the goal aimed
- Some of the variable loadings, among the smallests, are set to 0 thanks to a LASSO (L^1) penalty
- Associated variables are not taken into account when calculating the PCs



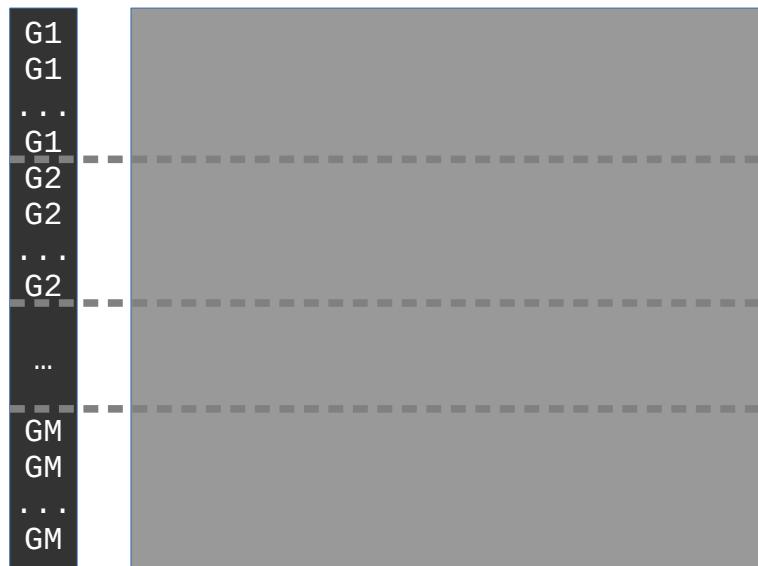
Vertical integration

A. Eslami, E.M Qannari, A. Kohler, S. Bougeard (2013). Analyses factorielles de données structurées en groupes d'individus. Journal de la SfdS, vol. 154(3). journal-sfds.fr/article/view/208

www.rocq.inria.fr/axis/modulad///sda11/HCSDA11-Qannari.PDF



- *Setting: the same variables measured on individuals portioned into several groups*
- *The same setting as in discriminant analysis **but** the main aim herein is to investigate the relationships among individuals within the various groups*



**Ask the
right
question!**

Vertical integration

How to investigate the relationships among individuals within the various groups?



Multivariate analysis of multi-group datasets

El Mostafa Qannari Aida Eslami
Achim Kohler Stéphanie Bougeard

- **Perform PCA on each group separately**
 - Too many parameters (stability and interpretation problems)
- **Perform PCA on the concatenated dataset**
 - The total variance recovered by the principal components mix up both the between and within groups variances
- **Multi-group PCA**
 - Perform PCA on the concatenated dataset **after centering by group**

Vertical integration: mgPCA

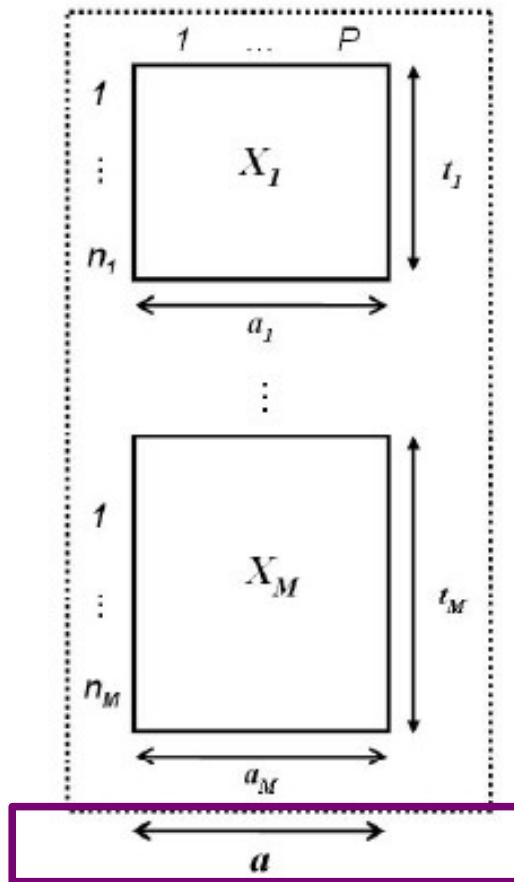
Oniris Institut National de l'Information Géographique et du Numérique
Sorbonne Université, AgroParisTech, INRAE, CNRS

anses

GENE

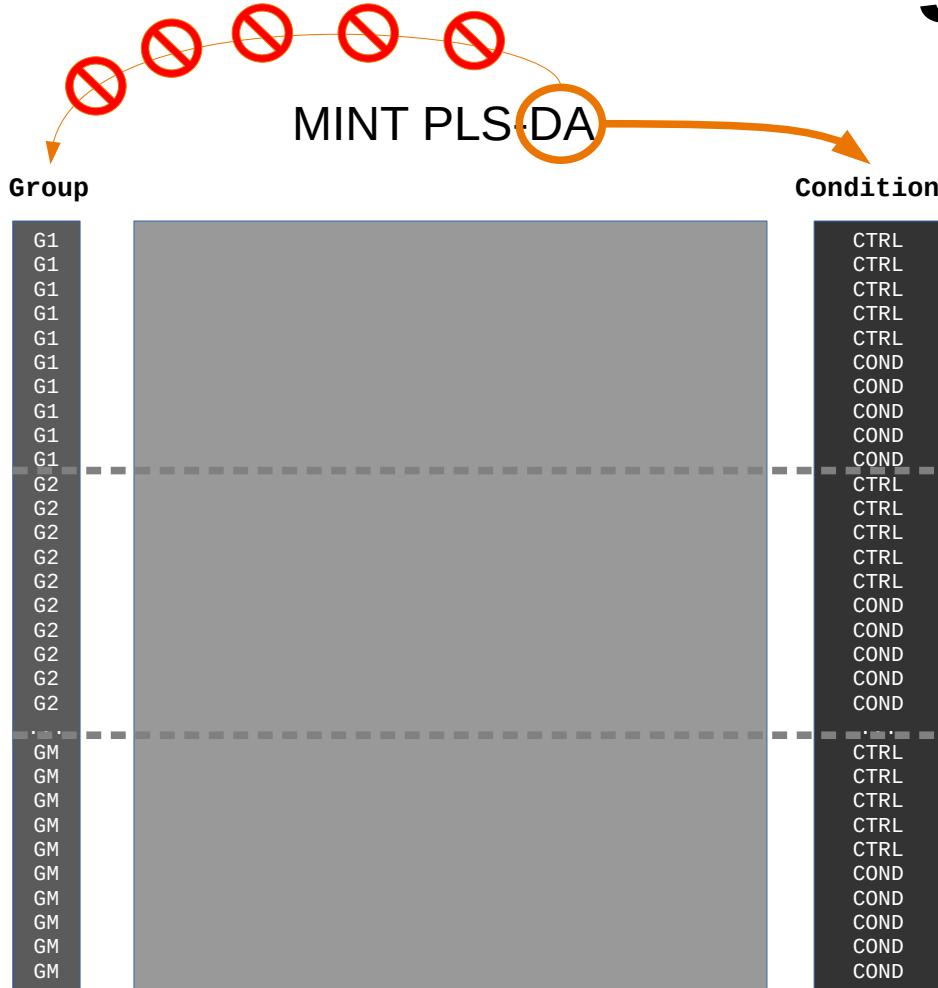
Multivariate analysis of multi-group datasets

El Mostafa Qannari Aida Eslami
Achim Kohler Stéphanie Bougeard



a: vector of common loadings
 → the same variable plot for every group

Vertical data integration

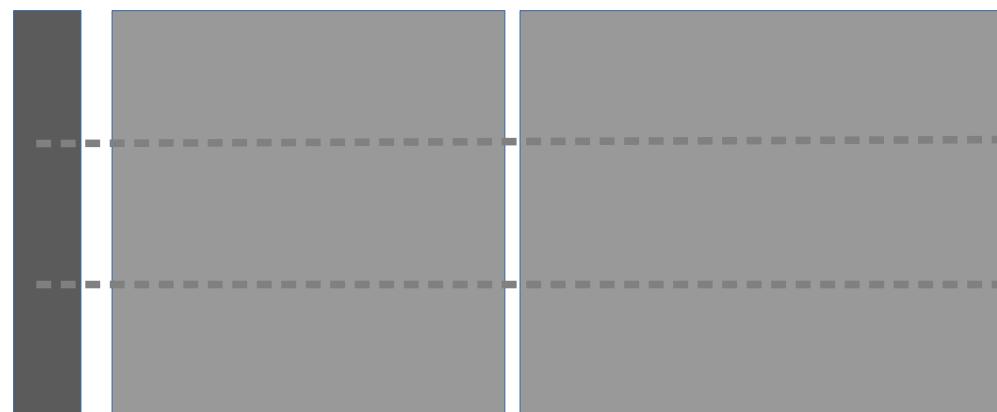


MINT: a multivariate integrative method to identify reproducible molecular signatures across independent experiments and platforms

Florian Rohart¹, Aida Eslami², Nicholas Matigian¹, Stéphanie Bougeard³ and Kim-Anh Lê Cao^{1*}

While PLS-DA ignores the data group structure inherent to each independent study, it can give satisfactory results when the between groups variance is smaller than the within group variance.

MINT PLS





Vertical data integration

MINT: a multivariate integrative method to identify reproducible molecular signatures across independent experiments and platforms

Florian Rohart¹, Aida Eslami², Nicholas Matigian¹, Stéphanie Boueard³ and Kim-Anh Lê Cao^{1*}

the component. For each dimension $h = 1, \dots, H$ PLS-DA seeks to maximize

$$\max_{\|a_h\|_2 = \|b_h\|_2 = 1} \text{cov}(X_h a_h, Y_h b_h), \quad (1)$$

For each dimension $h = 1, \dots, H$ the MINT algorithm seeks to maximize (m) group index

$$\max_{\|a_h\|_2 = \|b_h\|_2 = 1} \sum_{m=1}^M n_m \text{cov}(X_h^{(m)} a_h, Y_h^{(m)} b_h) + \lambda_h \|a_h\|_1,$$

a_h : vector of common loadings

In mgPLS, the PLS-components of each group are constraint to be built based on the same loading vectors in X and Y . These *global* loading vectors thus allow the samples from each group or study to be projected in the same common space spanned by the PLS-components.

We used a “Leave-One-Group-Out Cross-Validation (LOGOCV)”, which consists in performing CV where group or study m is left out only once $m = 1, \dots, M$. LOGOCV realistically reflects the true case scenario where prediction is performed on independent external studies based on a reproducible signature identified on the training set.

Examples

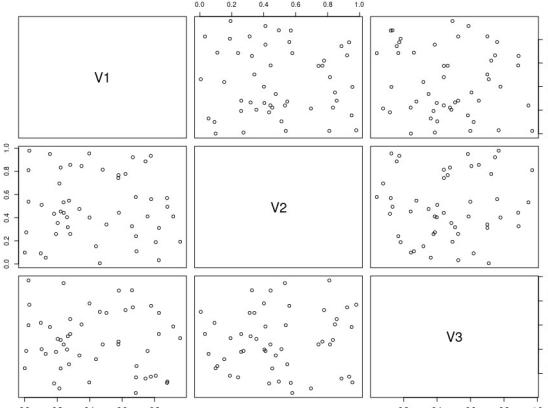
- Simulated toy examples for PCA, PLS-DA, PLS (2 blocks), multi-block PLS-DA
- `liver.toxicity` dataset (included in the package): practical session

PCA: simulated examples

Data set : 50 observations, 3 variables ($V1 - V2 - V3$)

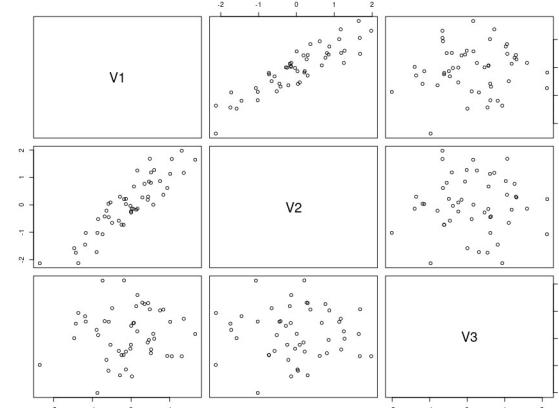
Case 1)

$\{V1\} - \{V2\} - \{V3\}$



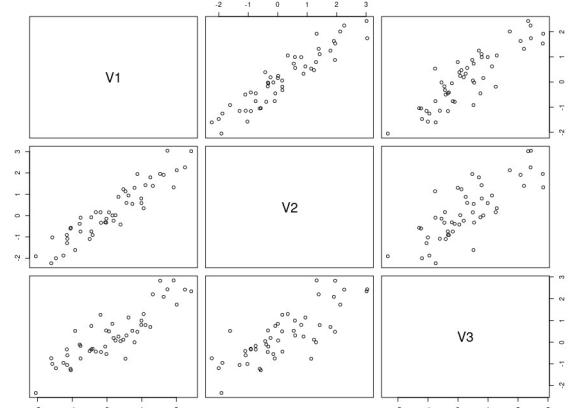
Case 2)

$\{V1 - V2\} - \{V3\}$



Case 3)

$\{V1 - V2 - V3\}$



Pearson Correlation matrices

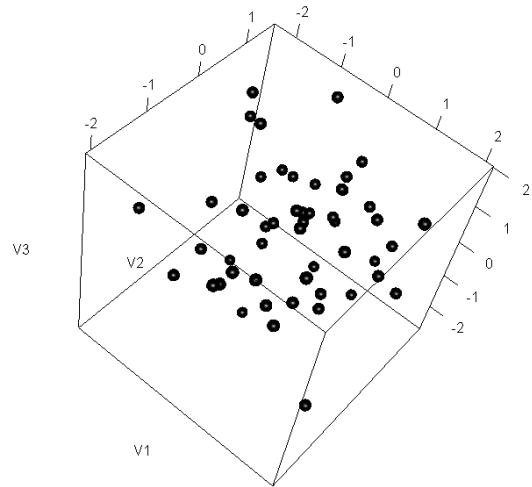
1)	V1	V2	V3
V1	1.00	-0.05	-0.12
V2	-0.05	1.00	0.06
V3	-0.12	0.06	1.00

2)	V1	V2	V3
V1	1.00	0.90	0.08
V2	0.90	1.00	-0.01
V3	0.08	-0.01	1.00

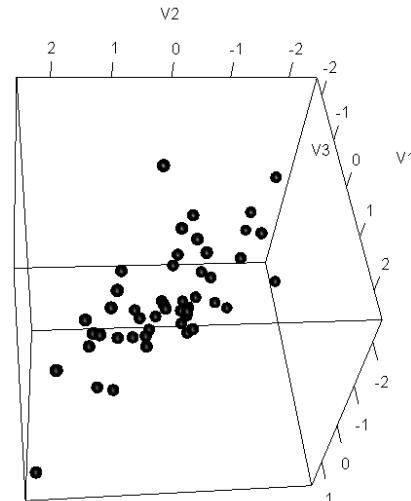
3)	V1	V2	V3
V1	1.00	0.93	0.87
V2	0.93	1.00	0.79
V3	0.87	0.79	1.00

PCA: simulated examples

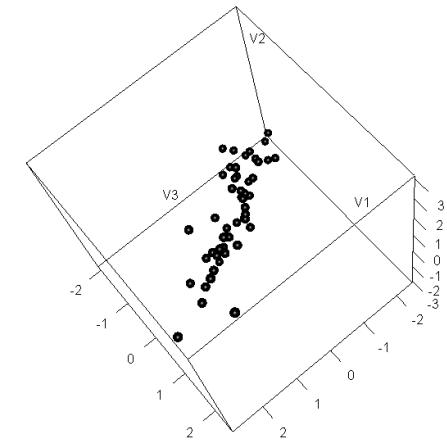
Case 1)



Case 2)



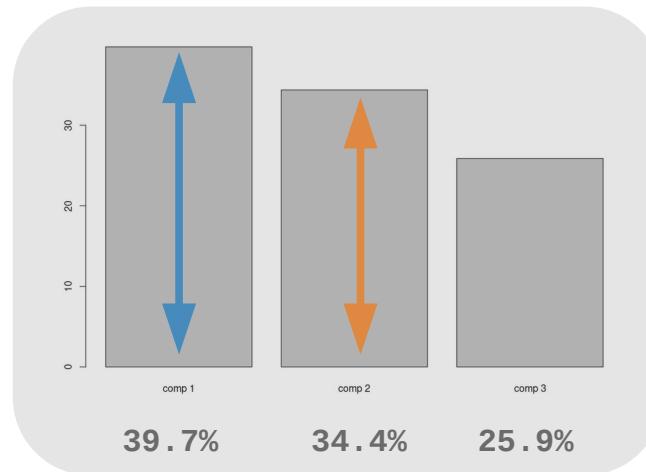
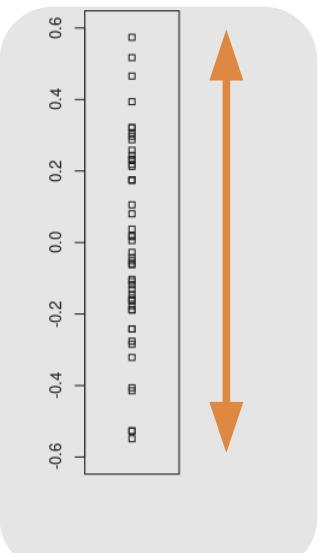
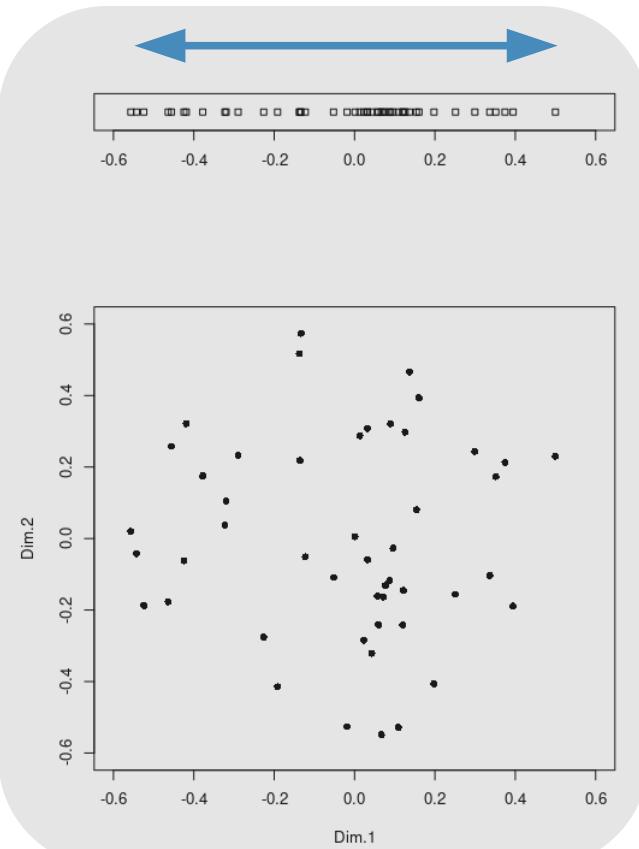
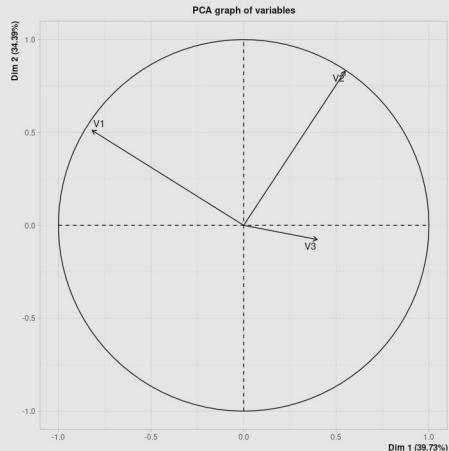
Case 3)



PCA: simulated examples

Loadings

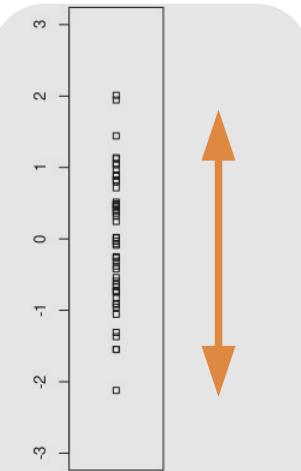
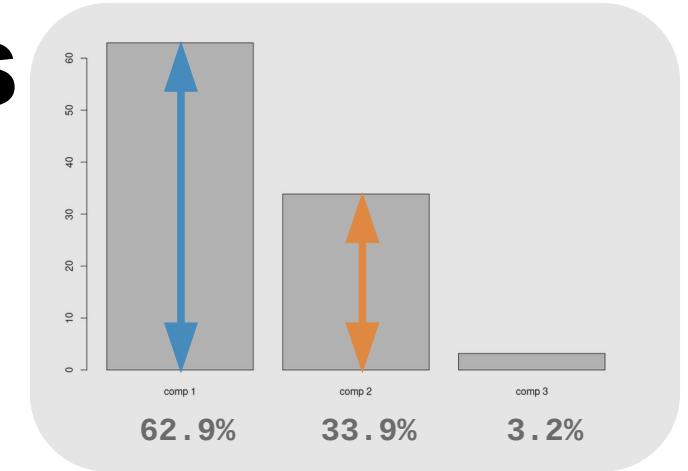
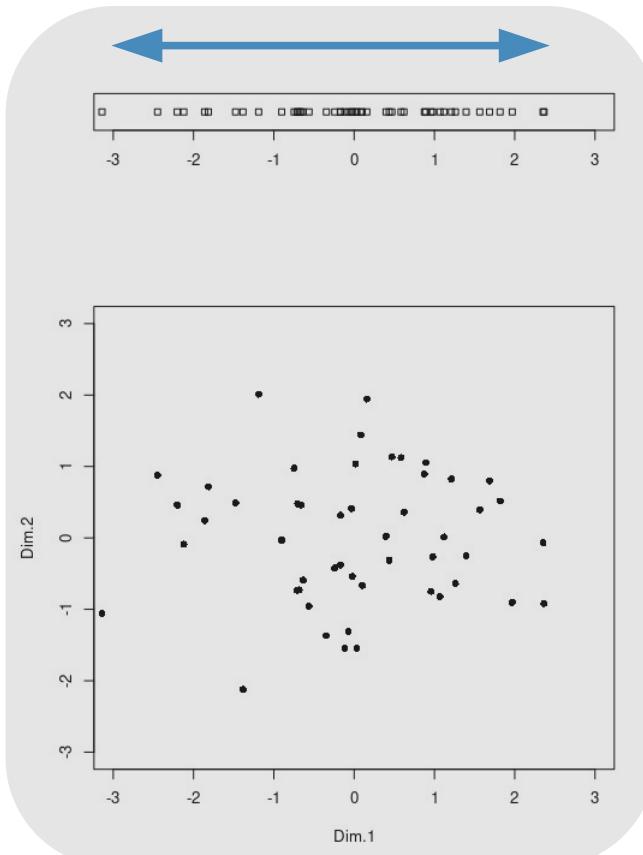
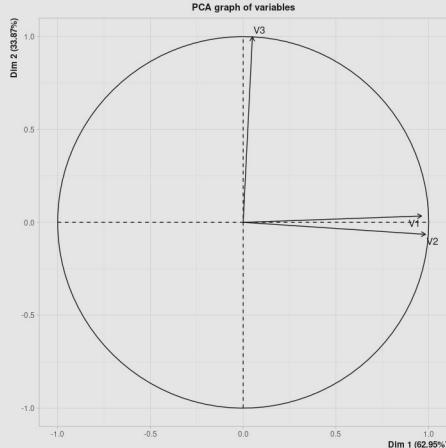
	Dim.1	Dim.2	Dim.3
V1	-0.23	0.14	0.07
V2	0.15	0.23	-0.03
V3	0.10	-0.02	0.22



PCA: simulated examples

Loadings

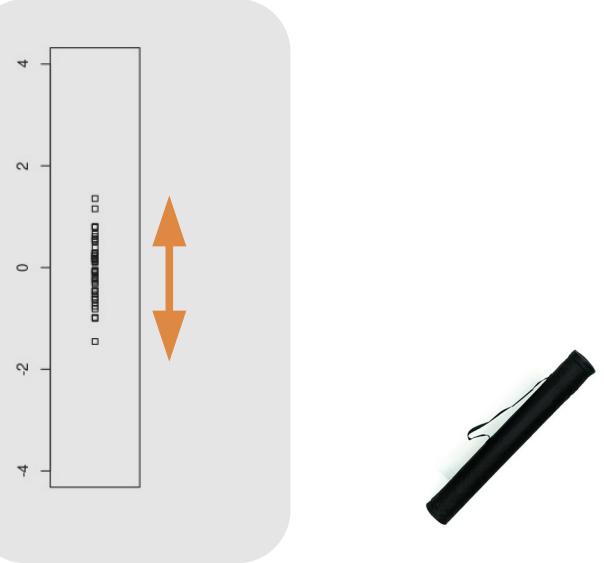
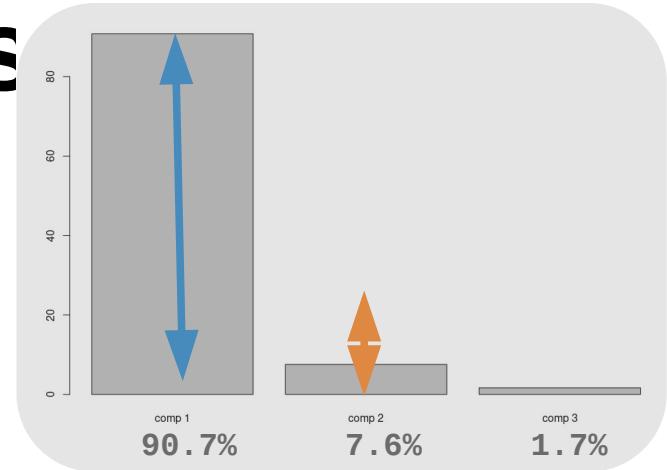
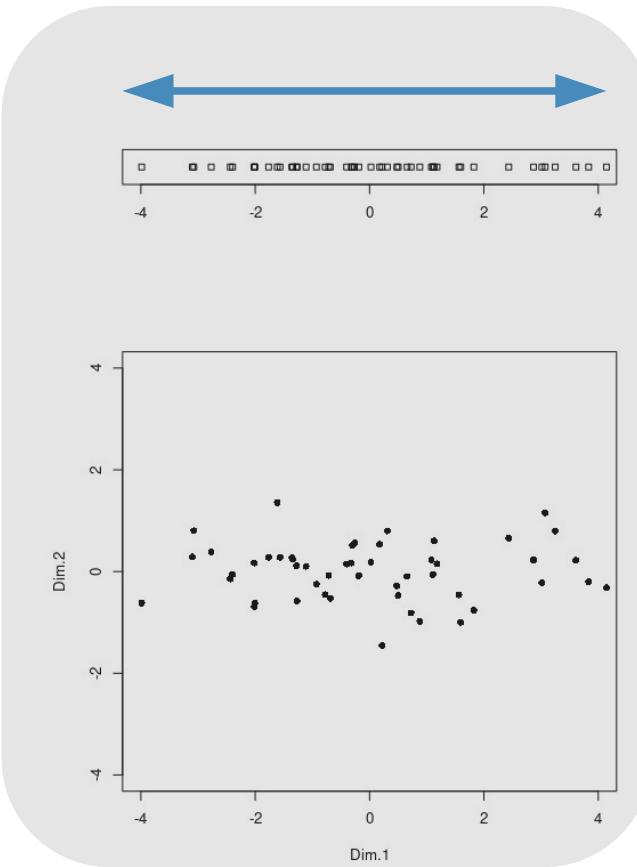
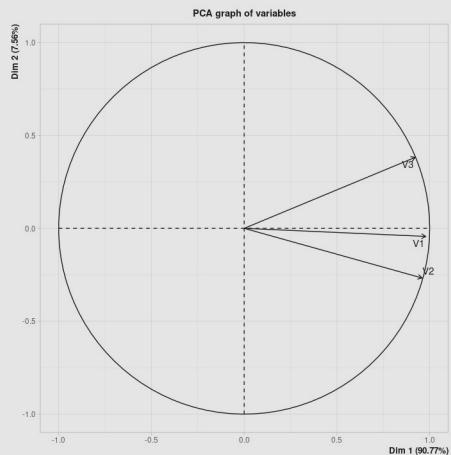
	Dim.1	Dim.2	Dim.3
V1	0.77	0.03	0.22
V2	0.97	-0.06	-0.17
V3	0.05	0.91	-0.02



PCA: simulated examples

Loadings

	Dim.1	Dim.2	Dim.3
V1	1.07	-0.05	0.22
V2	1.23	-0.34	-0.13
V3	1.07	0.44	-0.07



Discriminant analysis

Explore a data set composed of numerical variables and one categorical variable in order to separate the individuals based on their membership to the levels of the categorical variable

- Linear Discriminant Analysis (LDA): standard method (*underneath, same as PCA: matrix algebra*), needs more individuals than variables
- Projection to Latent Structure - Discriminant Analysis (PLS-DA): (*underneath, PLS algorithm*)



DA: simulated example

- 50 observations
- 4 variables :
 - 3 numerical: V1, V2, V3
 - 1 categorical: Group (A / B)

	V1	V2	V3
Mean	0.502	0.351	-0.076
Variance	17.28	8.44	1.49

- Bivariate analysis (Pearson correlation)

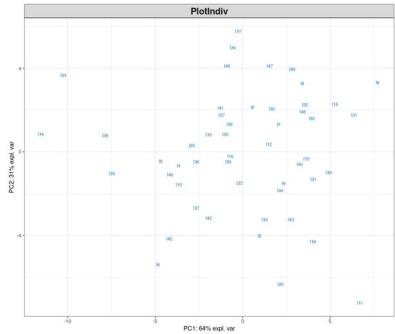
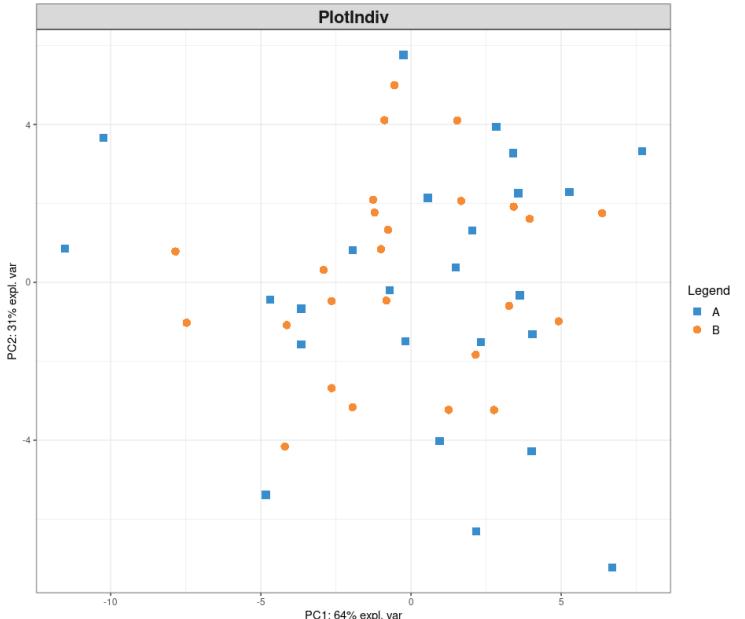
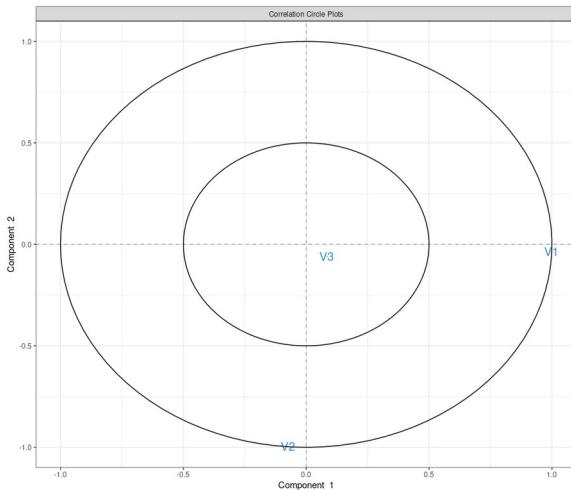
	V1	V2	V3
V1	1.000	-0.039	0.079
V2	-0.039	1.000	0.043
V3	0.079	0.043	1.000

V1	V2	V3	Group
2.88	1.71	1.19	A
-0.78	-1.33	-1.04	B
-0.82	-1.63	-1.96	B
5.62	-2.23	0.90	A
-0.30	-0.91	-1.26	B
-4.10	5.94	1.88	A
7.98	-3.39	1.08	A
-6.89	1.78	-0.85	B
3.48	3.48	-1.83	B
-3.11	2.07	1.72	A
2.44	-1.10	1.36	A
3.10	-3.76	0.95	A
6.79	-1.71	-0.97	B
-1.97	3.19	-1.15	B
-7.35	0.00	-1.02	B
-2.07	1.00	-1.77	B
2.97	6.52	0.91	A
-2.39	0.22	-1.34	B
-3.54	1.68	-1.38	B
-1.53	-0.41	1.84	A
-1.27	3.62	-0.41	B
3.69	-3.13	1.46	A
-3.13	1.19	0.69	A
-11.07	0.08	0.67	A
1.95	-0.12	0.77	A
-0.22	0.57	0.80	A
-0.28	0.87	-0.64	B
-0.51	-0.41	-1.29	B
0.92	-1.84	1.02	A
-0.08	-5.41	0.84	A
2.09	-1.77	-1.16	B
2.78	2.10	-1.23	B
1.65	4.30	0.67	A
-9.95	-2.80	0.91	A
1.82	-3.83	0.04	B
-0.56	-3.67	-1.64	B
-0.27	-4.57	-1.76	B
3.91	-2.13	1.67	A
5.50	1.11	-1.33	B
1.94	3.53	-0.73	B
4.72	4.40	0.86	A
7.56	7.20	0.68	A
3.84	-1.72	-0.99	B
3.82	0.80	-1.03	B
-4.18	1.02	0.54	A
-3.44	4.76	-1.56	B
4.41	-1.42	-1.82	B
4.59	1.43	0.75	A
4.11	0.46	1.25	A
0.37	1.83	0.89	A

DA: simulated example

- Multivariate **unsupervised** analysis (PCA)

Loadings	PC1	PC2	PC3
V1	0.998	-0.052	0.026
V2	-0.053	-0.998	0.023
V3	0.025	-0.025	-0.999

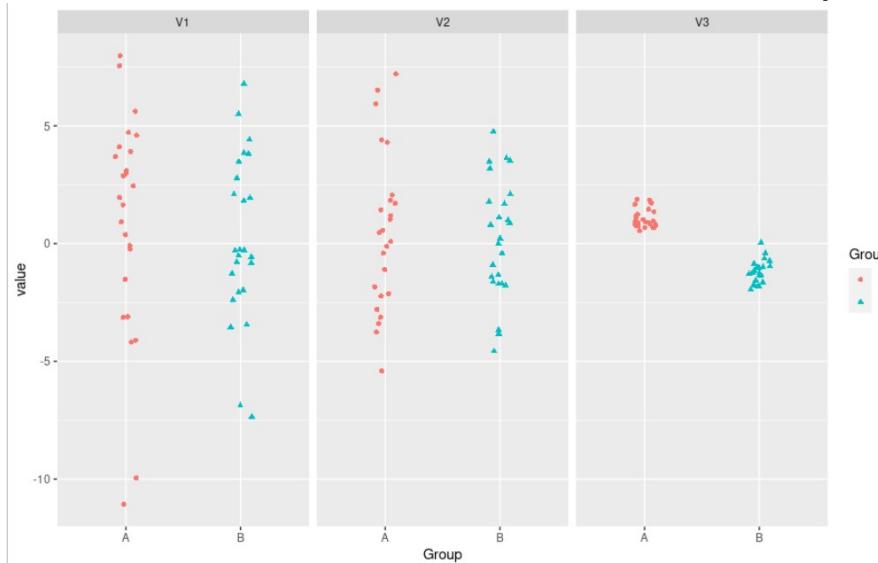


DA: simulated example

- Linear Discriminant Analysis

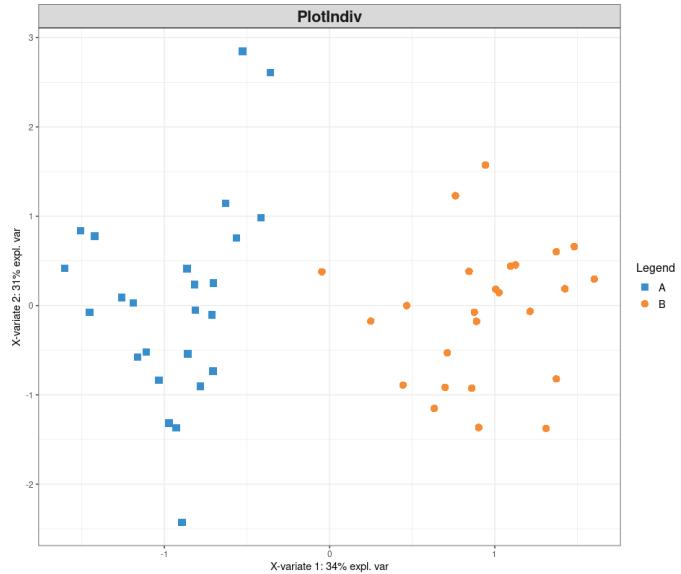
Loadings	LD1
V1	-0.007
V2	-0.011
V3	-2.295

V3 is highly involved in the discrimination of the two groups (even with a small variance)



- PLS Discriminant Analysis

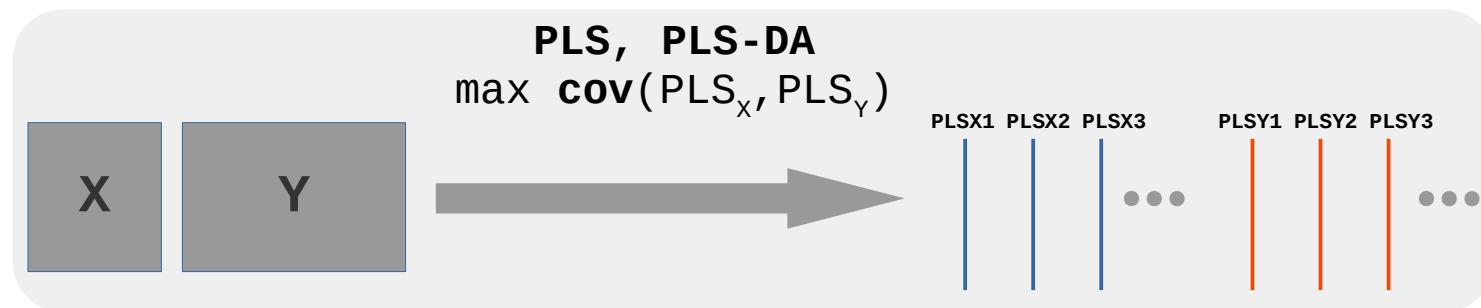
Loadings	PLSDA1	PLSDA2	PLSDA3
V1	-0.088	-0.888	-0.452
V2	-0.053	-0.449	0.892
V3	-0.995	0.103	-0.008



Two-block integration

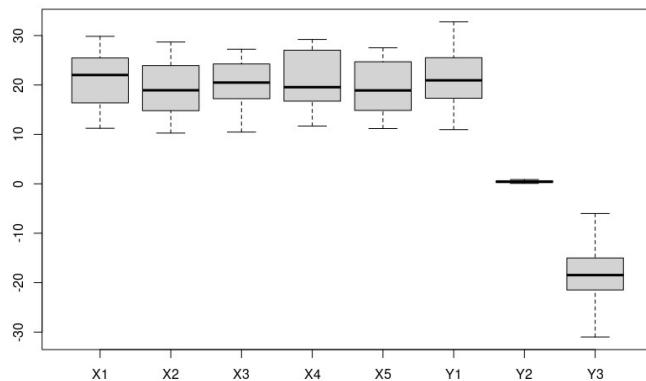
Unravel the relationships between two types of variables measured on the same matching samples

- Understand the correlation/covariance structure between two data sets
- Select co-regulated biological entities across samples
- Methods: Projection to Latent Structures (**PLS, maximize covariance**)



Two-block: simulated example

- 20 observations
- 2 sets of numerical variables:
 - X: 5 variables
 - Y: 3 variables
- Univariate analysis

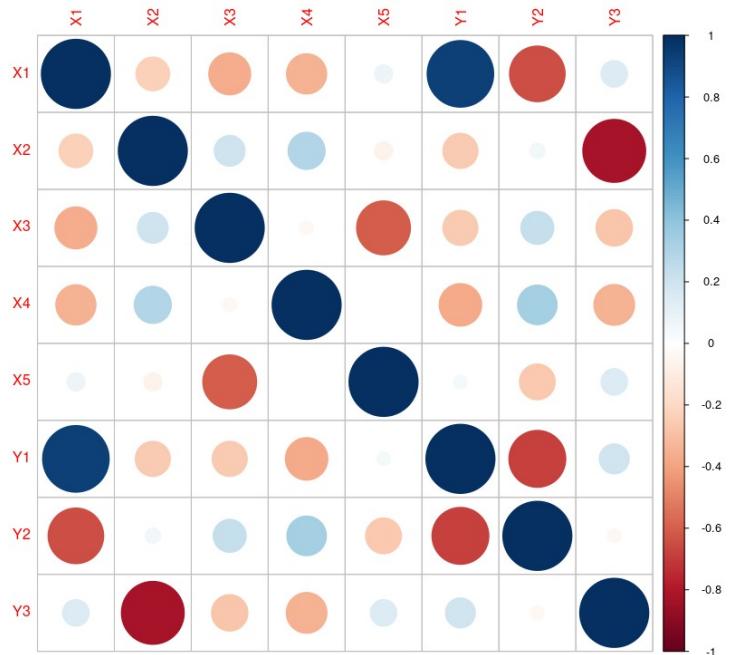


X1	X2	X3	X4	X5	Y1	Y2	Y3
15.3	28.7	26.4	28.3	18.7	16.1	0.7	-31.0
17.4	14.2	22.9	15.9	24.3	16.2	0.6	-14.8
21.5	23.0	25.7	19.2	18.0	22.1	0.2	-18.3
28.2	12.5	21.1	16.6	16.5	25.9	0.3	-18.6
14.0	15.3	20.6	23.0	25.1	16.9	0.7	-13.0
28.0	17.7	25.8	15.2	14.1	31.9	0.5	-16.4
28.9	10.3	10.5	19.6	24.2	28.2	0.2	-6.0
23.2	17.6	19.5	25.3	12.4	21.1	0.7	-18.9
22.6	27.4	24.6	11.7	14.9	23.7	0.1	-25.9
11.2	16.8	23.9	27.5	12.9	11.0	0.9	-15.7
14.1	19.6	19.6	16.8	14.8	18.9	0.6	-21.8
13.5	22.0	27.2	26.8	11.2	13.5	0.6	-17.2
23.7	19.9	18.8	16.9	22.8	25.1	0.3	-15.2
17.7	13.7	14.9	16.7	27.5	17.7	0.5	-10.9
25.4	26.5	11.4	19.5	25.6	23.9	0.5	-20.2
20.0	23.4	12.0	27.8	25.9	20.3	0.2	-21.1
24.4	25.9	16.3	27.3	19.1	20.7	0.5	-31.0
29.8	12.2	20.4	17.8	18.2	32.8	0.1	-14.5
17.6	24.5	23.2	25.5	26.2	17.9	0.3	-29.4
25.5	18.2	18.1	29.2	22.1	29.9	0.2	-20.1

Two-block: simulated example

- Bivariate analysis

	X1	X2	X3	X4	X5	Y1	Y2	Y3
X1	1.00	-0.24	-0.37	-0.34	0.07	0.93	-0.65	0.15
X2	-0.24	1.00	0.20	0.29	-0.07	-0.26	0.05	-0.83
X3	-0.37	0.20	1.00	-0.04	-0.61	-0.26	0.23	-0.28
X4	-0.34	0.29	-0.04	1.00	0.00	-0.38	0.33	-0.35
X5	0.07	-0.07	-0.61	0.00	1.00	0.04	-0.27	0.15
Y1	0.93	-0.26	-0.26	-0.38	0.04	1.00	-0.68	0.19
Y2	-0.65	0.05	0.23	0.33	-0.27	-0.68	1.00	-0.04
Y3	0.15	-0.83	-0.28	-0.35	0.15	0.19	-0.04	1.00



Main effects: (X1, Y1) positively correlated, (X1, Y2) and (X2, Y3) negatively correlated

Two-block: simulated example

Projection to Latent Structure

Loadings
(2 sets, one for
each dataset)

	CCA.X1	CCA.X2	CCA.X3
X1	-0.73	-0.58	-0.27
X2	0.42	-0.80	0.35
X3	0.30	-0.13	-0.49
X4	0.42	-0.08	-0.17
X5	-0.17	0.06	0.73

	CCA.Y1	CCA.Y2	CCA.Y3
Y1	-0.71	-0.48	-0.88
Y2	0.51	0.45	-0.24
Y3	-0.49	0.75	-0.41



What is underneath?
An iterative algorithm

Two-block: simulated example

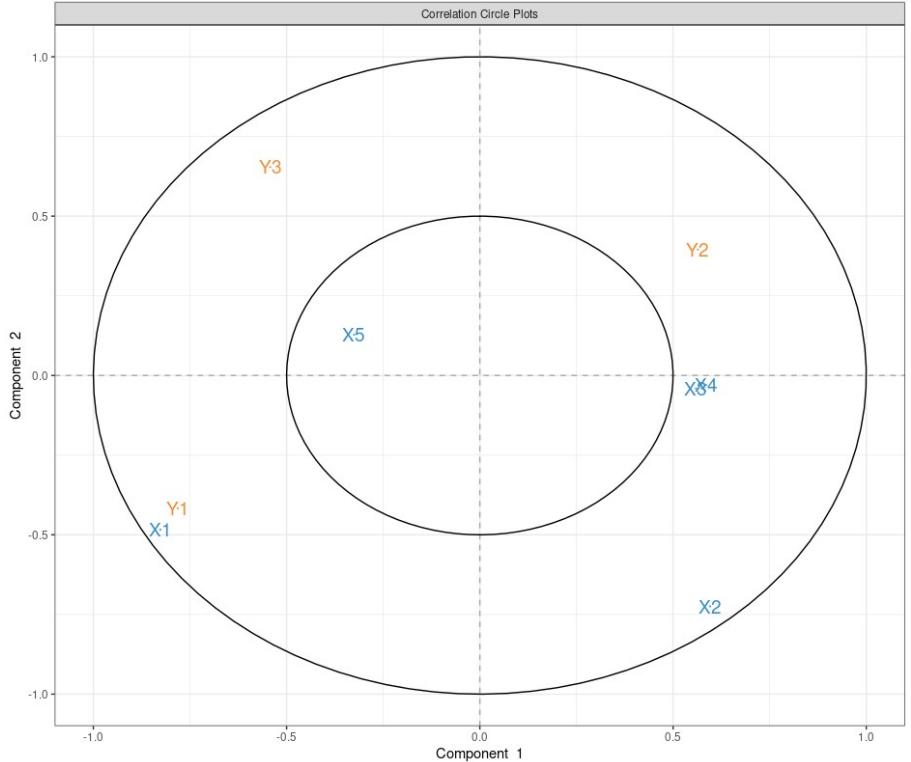
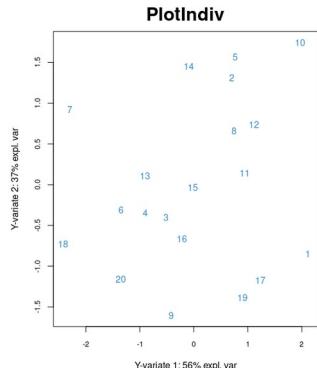
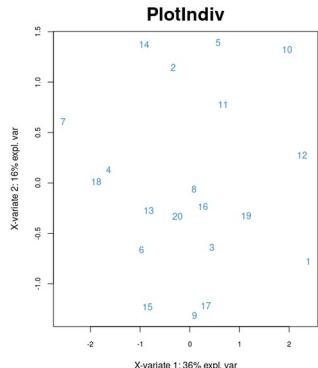
PLS.X1	PLS.X2	PLS.X3	PLS.Y1	PLS.Y2	PLS.Y3
2.38	-0.78	0.21	2.11	-0.85	-0.07
-0.33	1.15	0.33	0.70	1.31	0.55
0.44	-0.64	-0.44	-0.52	-0.40	-0.18
-1.64	0.13	-1.39	-0.90	-0.35	0.49
0.57	1.39	0.81	0.76	1.57	-0.32
-0.97	-0.66	-1.71	-1.35	-0.31	-1.12
-2.55	0.61	0.32	-2.29	0.92	-0.02
0.09	-0.06	-1.28	0.75	0.66	-0.30
0.10	-1.32	-0.36	-0.42	-1.61	0.42
1.96	1.33	-0.93	1.97	1.75	-0.23
0.67	0.78	-0.05	0.94	0.14	0.00
2.26	0.28	-1.21	1.11	0.74	-0.37
-0.82	-0.27	0.49	-0.91	0.11	-0.15
-0.92	1.37	1.41	-0.09	1.45	0.58
-0.85	-1.23	1.83	-0.02	-0.03	0.19
0.26	-0.23	1.78	-0.22	-0.66	0.43
0.33	-1.22	0.39	1.24	-1.17	0.61
-1.88	0.01	-1.26	-2.42	-0.73	-0.45
1.13	-0.33	1.10	0.90	-1.38	0.70
-0.25	-0.33	-0.05	-1.35	-1.16	-0.77

$\text{cor}(\text{PLS.X1}, \text{PLS.Y1}) = 0.86$
 $\text{cov}(\text{PLS.X1}, \text{PLS.Y1}) = 1.46$

Two-block: simulated example

PLS.X1	PLS.X2	PLS.X3
2.38	-0.78	0.21
-0.33	1.15	0.33
0.44	-0.64	-0.44
-1.64	0.13	-1.39
0.57	1.39	0.81
-0.97	-0.66	-1.71
-2.55	0.61	0.32
0.09	-0.06	-1.28
0.10	-1.32	-0.36
1.96	1.33	-0.93
0.67	0.78	-0.05
2.26	0.28	-1.21
-0.82	-0.27	0.49
-0.92	1.37	1.41
-0.85	-1.23	1.83
0.26	-0.23	1.78
0.33	-1.22	0.39
-1.88	0.01	-1.26
1.13	-0.33	1.10
-0.25	-0.33	-0.05

PLS.Y1	PLS.Y2	PLS.Y3
2.11	-0.85	-0.07
0.70	1.31	0.55
-0.52	-0.40	-0.18
-0.90	-0.35	0.49
0.76	1.57	-0.32
-1.35	-0.31	-1.12
-2.29	0.92	-0.02
0.75	0.66	-0.30
-0.42	-1.61	0.42
1.97	1.75	-0.23
0.94	0.14	0.00
1.11	0.74	-0.37
-0.91	0.11	-0.15
-0.09	1.45	0.58
-0.02	-0.03	0.19
-0.22	-0.66	0.43
1.24	-1.17	0.61
-2.42	-0.73	-0.45
0.90	-1.38	0.70
-1.35	-1.16	-0.77

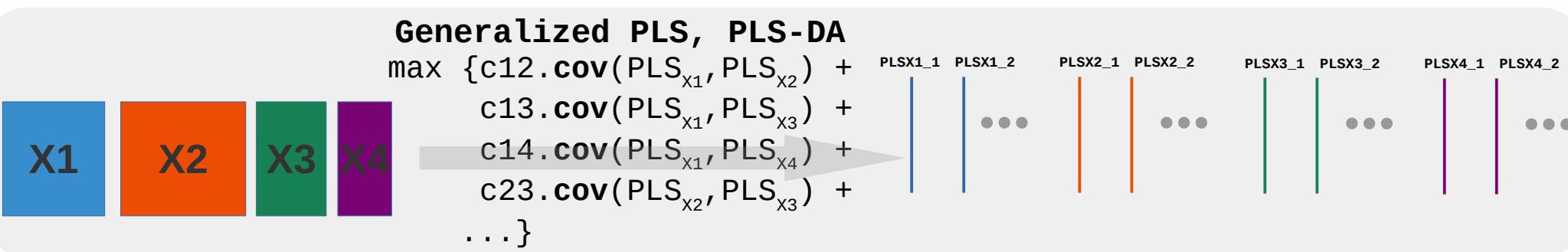


Variable plot clearly highlights the correlation structure between the two datasets: (X1, Y1) positively correlated, (X1, Y2) and (X2, Y3) negatively correlated

Multi-block integration

Unravel the relationships between **more than two** types of variables measured on the same matching samples

- Understand the relationships structure between several data sets
- Select co-regulated biological entities across samples
- Method: Multi-block PLS



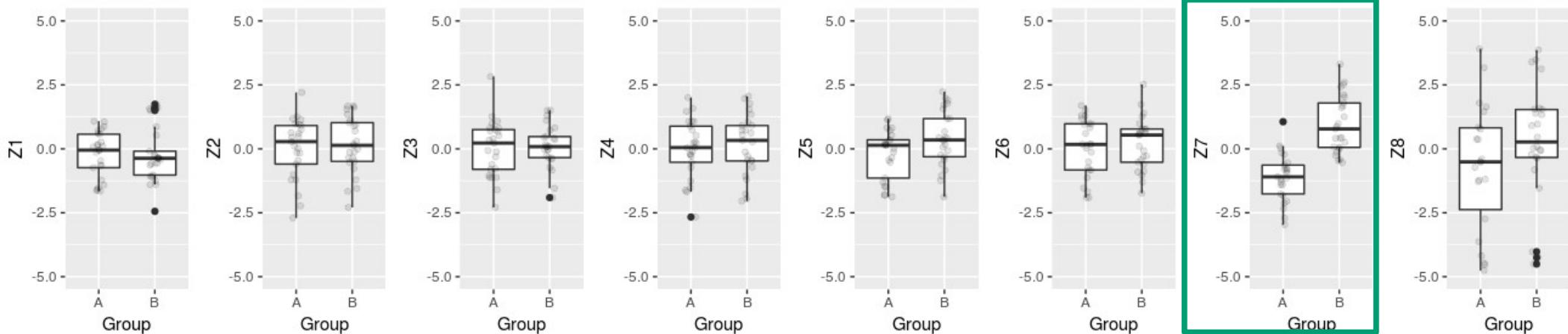
Multi-block: simulated example

- 50 observations
- 3 sets of numerical variables:
 - X: 5 variables
 - Y: 3 variables
 - Z: 8 variables
- 1 categorical variable Group A/B

Correlation matrix
between X, Y and
Z variables

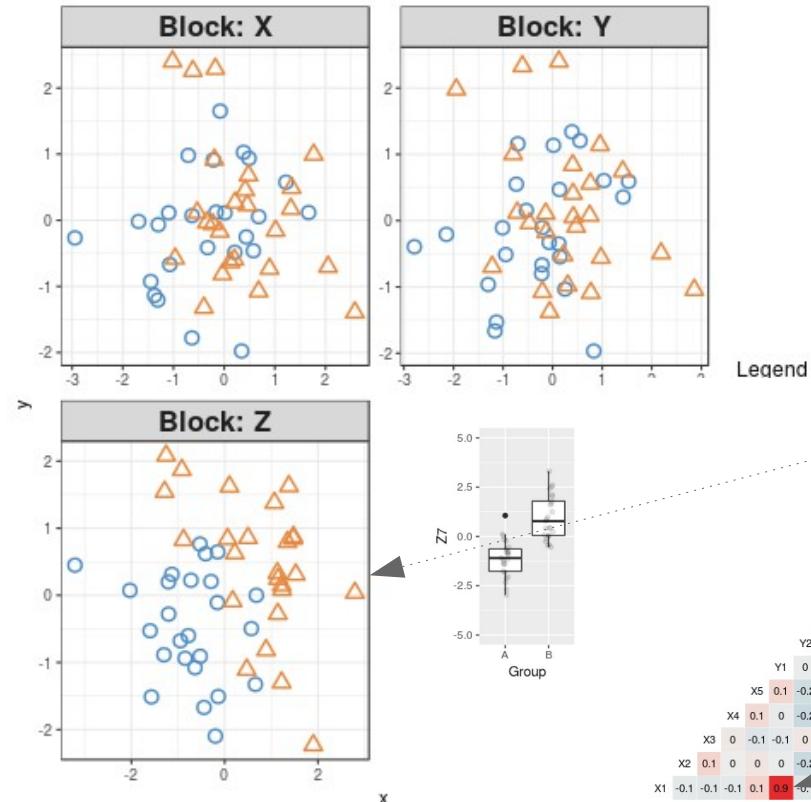
								Z8
								Z7 0.1
								Z6 -0.2 0
								Z5 0.2 0.1 0.2
								Z4 0.2 -0.1 0.1 0.2
								Z3 -0.2 0 -0.2 0.3 0.1
								Z2 -0.1 -0.2 -0.1 0.1 0.2 0.1
								Z1 0 0 0 0.1 0.1 -0.1 -0.1
								Y3 -0.1 0.1 0 0.2 0.2 0.1 0.2 1
								Y2 0.2 0 -0.1 -0.1 0 0.1 -0.1 0 0.2
								Y1 0 0 -0.1 0 0.1 0 -0.2 0.3 -0.3 0
								X5 0.1 -0.2 -0.1 0.2 0 0 0.1 0 0.1 0 -0.1
								X4 0.1 0 -0.2 0.1 -0.2 0.1 0.1 0.2 0 -0.1 0.3 0
								X3 0 -0.1 -0.1 0 0.7 -0.2 0 0.2 0 0.2 0.1 0.2 0.7
								X2 0.1 0 0 0 -0.2 -0.6 -0.1 0.2 -0.2 0 0.1 0 0 -0.6
								X1 -0.1 -0.1 -0.1 0.1 0.9 -0.1 0 0 -0.1 0.1 -0.1 -0.2 0.3 -0.4 0

Boxplot of the variables Z according to the group

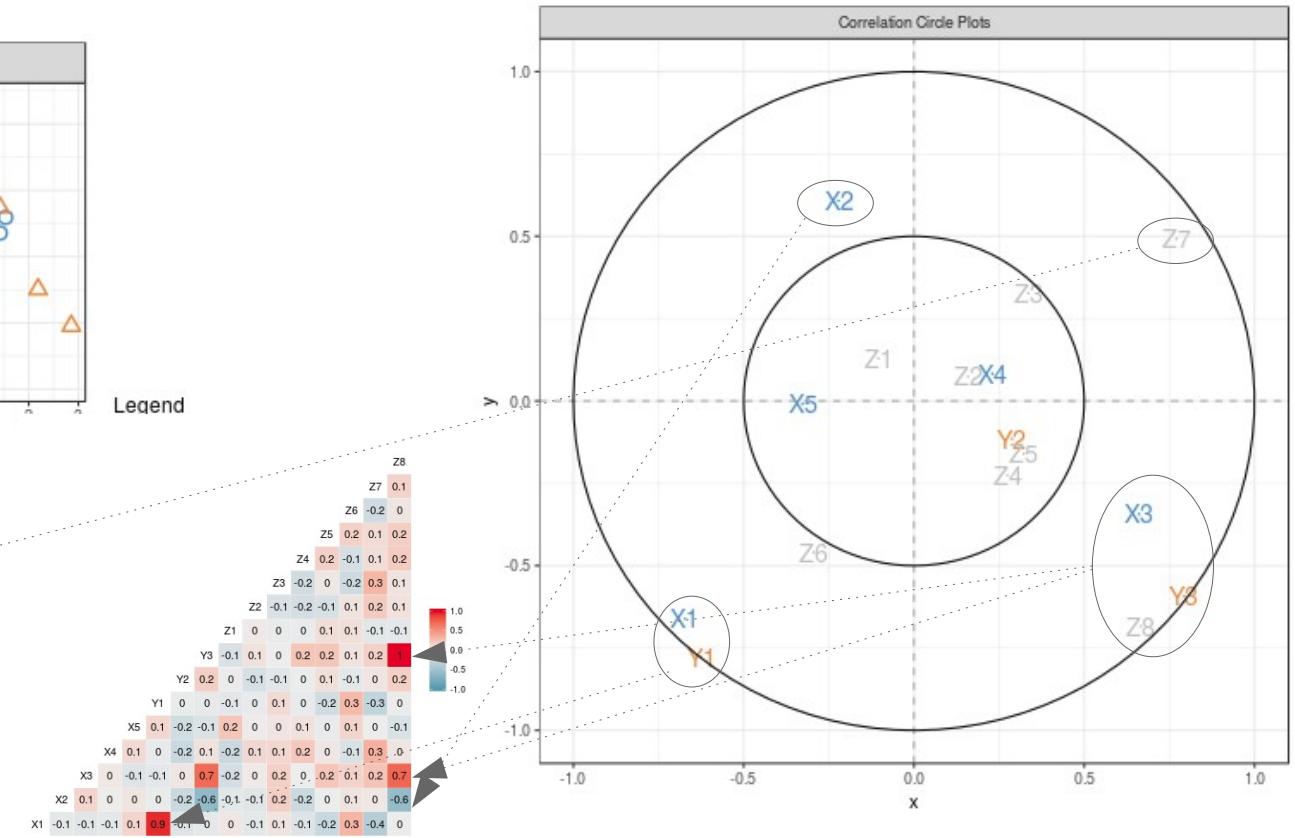


Multi-block: simulated example

Three individual plots



Variable plot



Take home message

- Practice on your own data! The best way to understand what a method has to tell you.
- Do not bypass the elementary analyses (univariate, bivariate, multivariate single data set).
- Address problems explicitly formulated: “I want to integrate my data” is not a problem explicitly formulated.
- Clearly identify supervised and unsupervised questions and the methods to use. “PCA is not a good method, I can’t see my clusters...” reveals a misunderstanding of PCA.

What about microbiome data?

Lê Cao KA, Costello ME, Lakis VA, Bartolo F, Chua XY, et al. (2016)
MixMC: A Multivariate Statistical Framework to Gain Insight into Microbial Communities.
PLOS ONE 11(8): e0160169. <https://doi.org/10.1371/journal.pone.0160169>

- mixMC: pipeline set up for microbial communities, using some of the standards methods in mixOmics but with a bit of tweaking
- due to the sparse and compositional nature (represent proportions or relative information) of microbiome data, there are specific pre-processing steps which need to be undergone in order to avoid spurious results
- Option 1: some of the functions (`pca()`, `plsda()`) include the argument `logratio = 'CLR'`
- Option 2: use the `logratio.transfo()` function

From the mixOmics newsletter (2023/05/15)

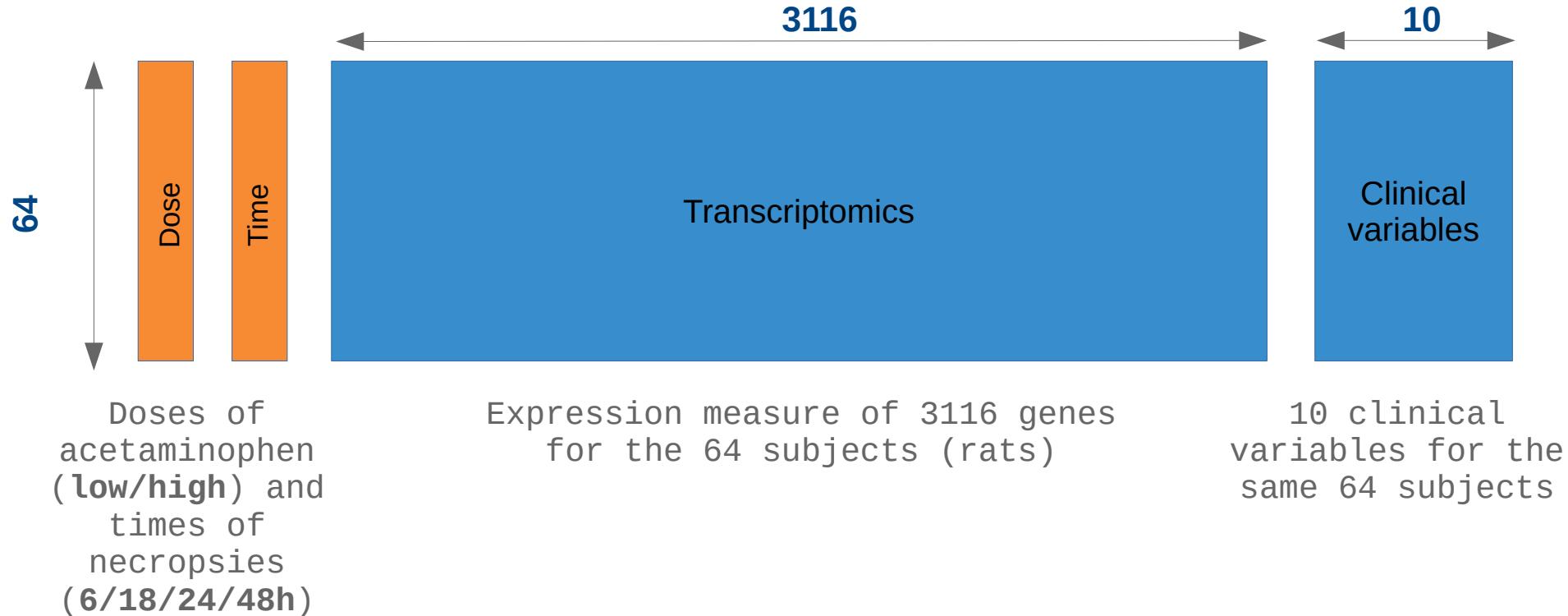
A new recording about time-course multi-omics integration is now available at this page:
mixomics.org/time-course-integration/

We are still working actively in this area, especially for **microbiome data**.

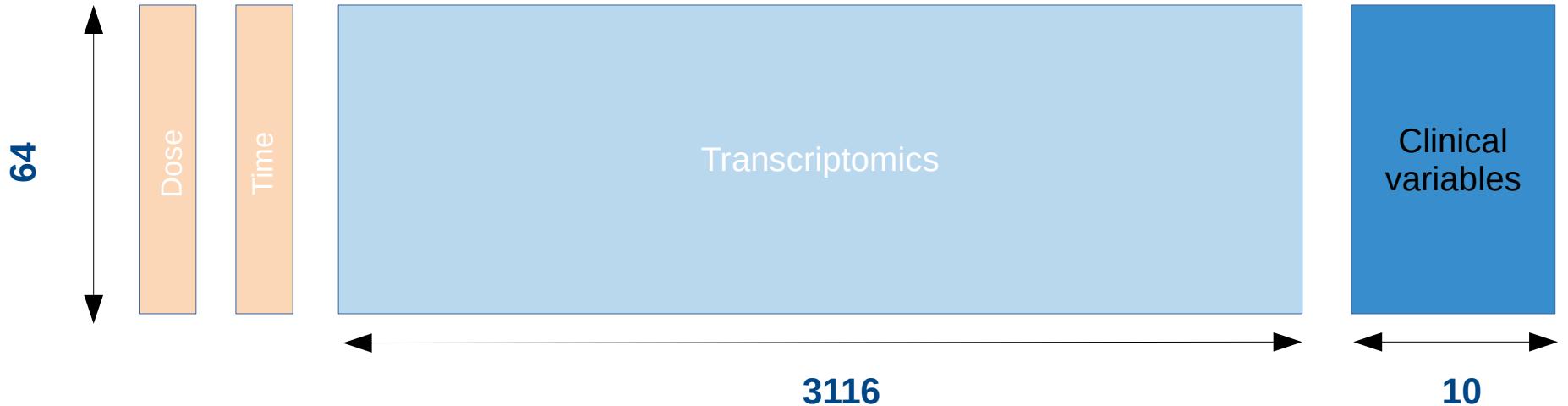
Examples

Bushel, P.R., Wolfinger, R.D. & Gibson, G. Simultaneous clustering of gene expression data with clinical chemistry and pathological evaluations reveals phenotypic prototypes. BMC Syst Biol 1, 15 (2007). <https://doi.org/10.1186/1752-0509-1-15>

```
R> library(mixOmics)  
R> data(liver.toxicity)  
R> help(liver.toxicity)
```

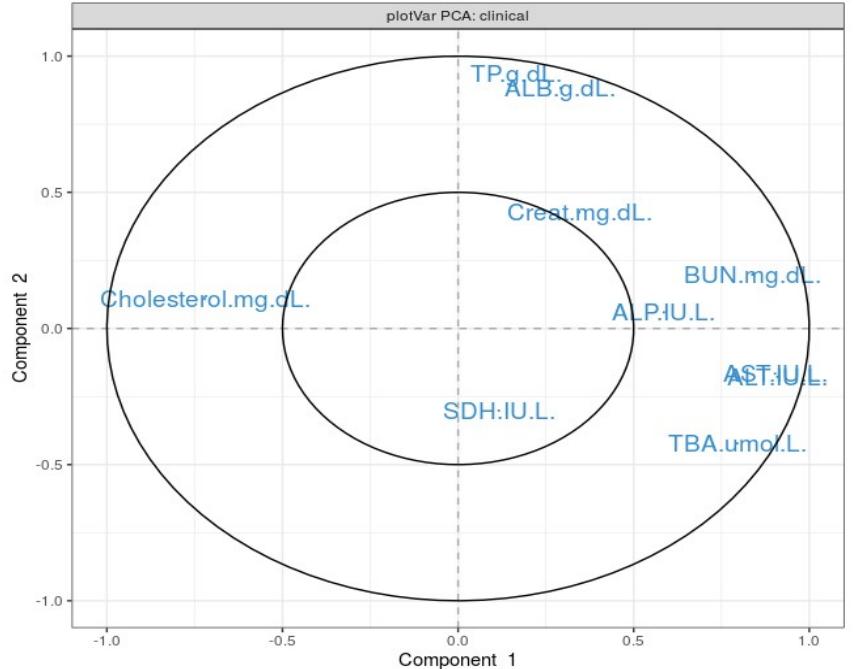
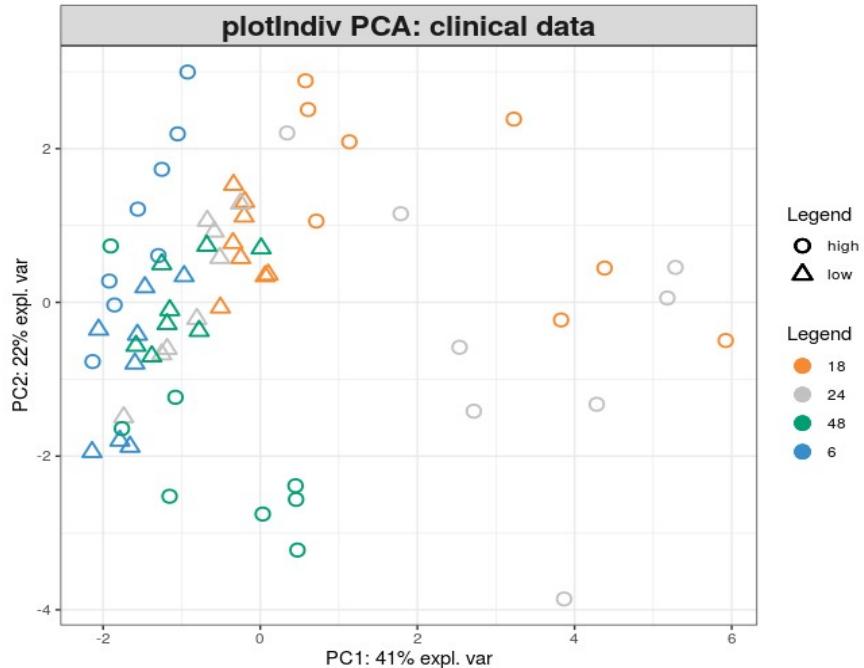


Explore one data set



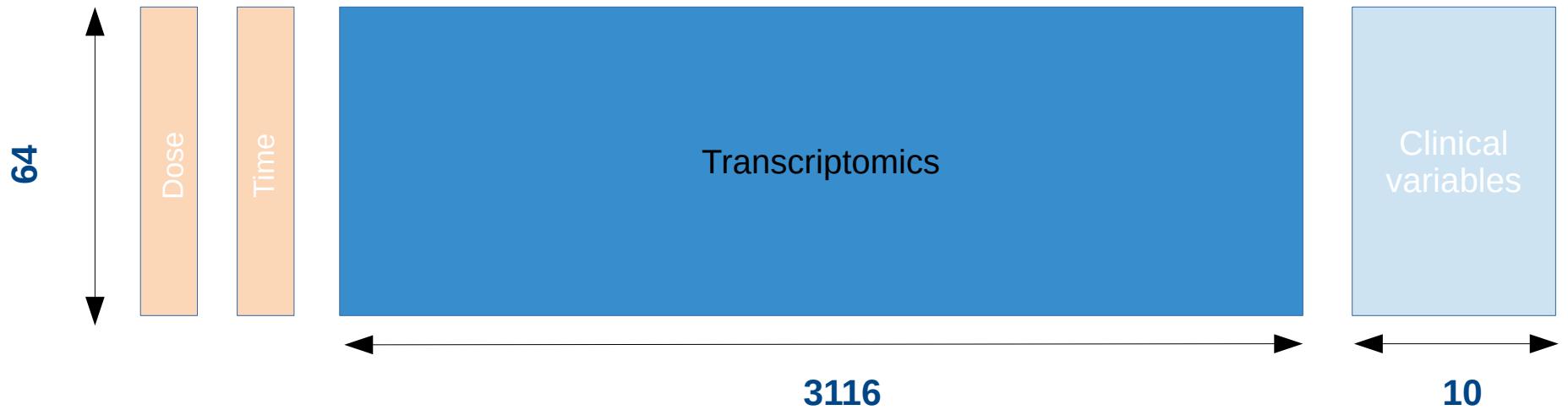
Question: based on clinical data, do we naturally observe clusters of samples which correspond to the different dose or exposure treatments?

PCA on clinical variables



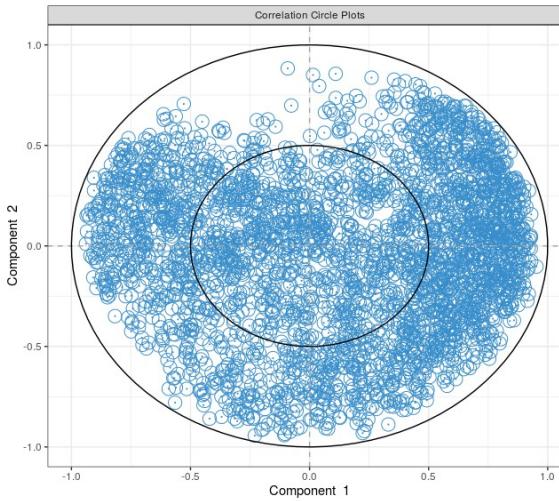
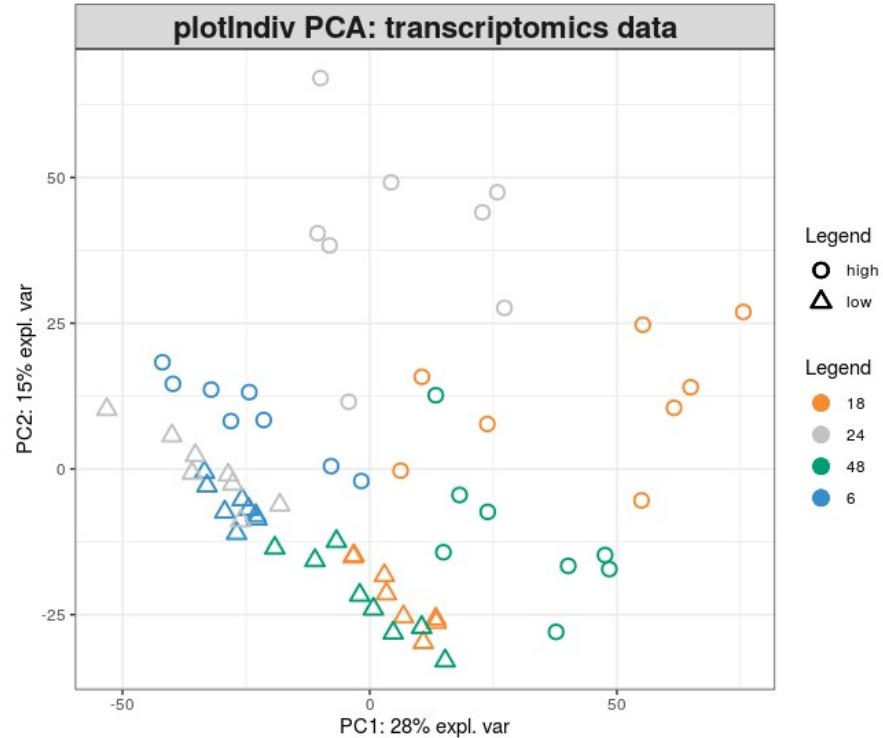
Answer: SO, SO...

Explore another data set



Question: based on transcriptomics data, do we naturally observe clusters of samples which correspond to the different dose or exposure treatments?

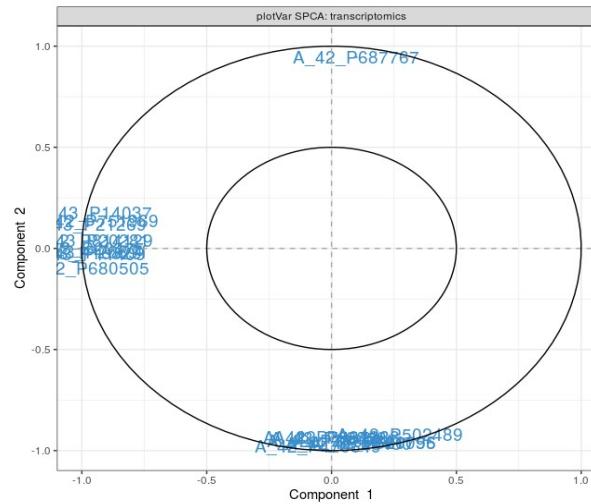
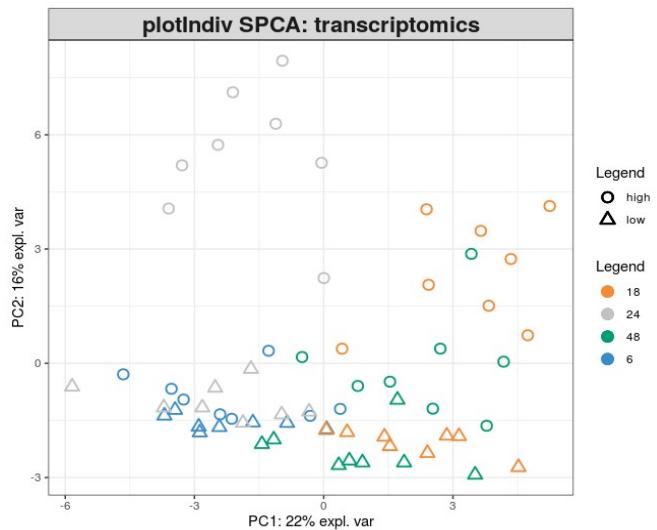
PCA on transcriptomics data



Answer: dose effect appears clearly as well as trends in time effect...

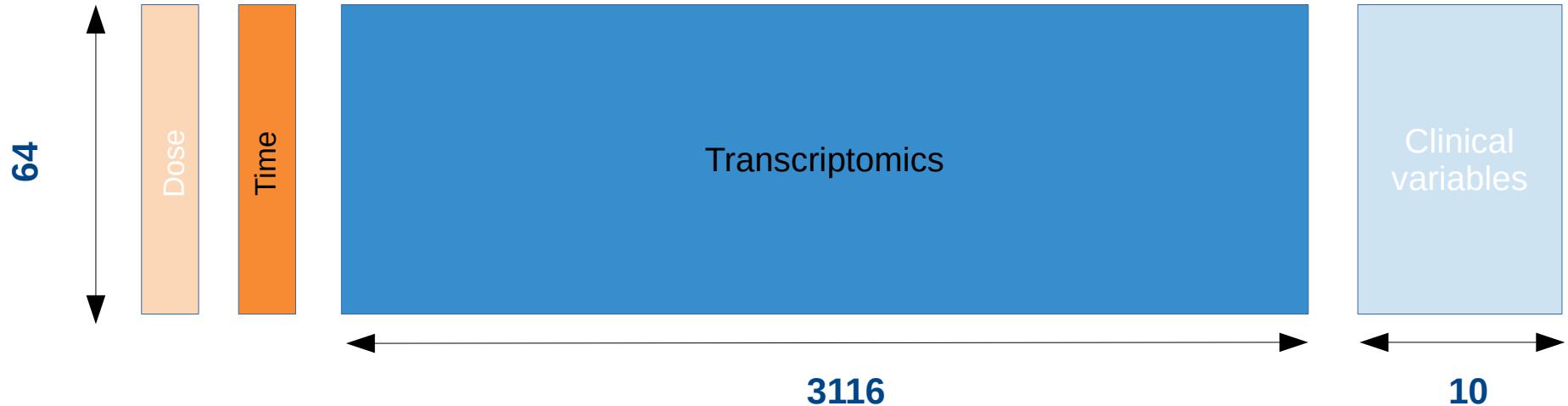
Too many genes? Sparse PCA

Question: based on transcriptomics data, do we naturally observe clusters of samples which correspond to the different doses or exposure treatments **when we select some genes highly involved in the variability of the data?**



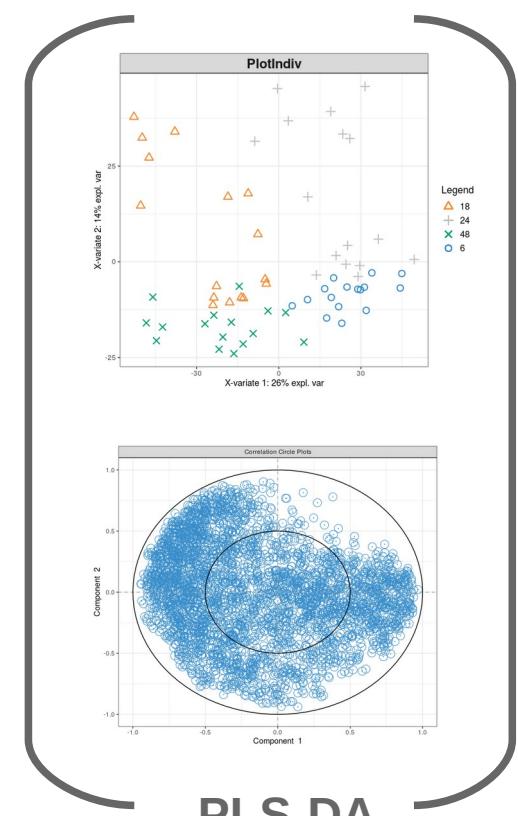
Answer: behaviour roughly similar when considering every gene or not.

Supervised analysis: transcriptomics / time

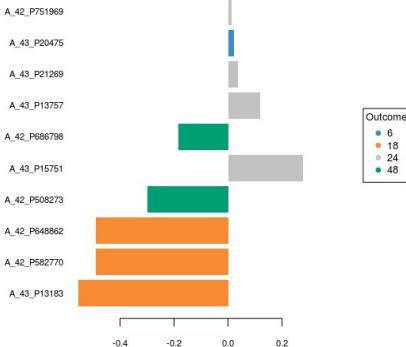


Question: Based on transcriptomics data, can we identify a molecular signature that characterizes the different treatment times?

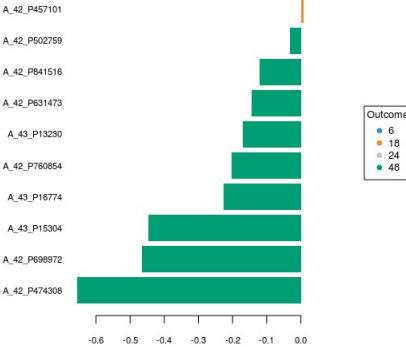
(S)PLS-DA transcript. / time



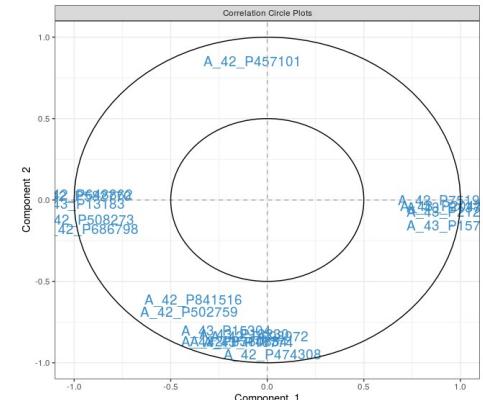
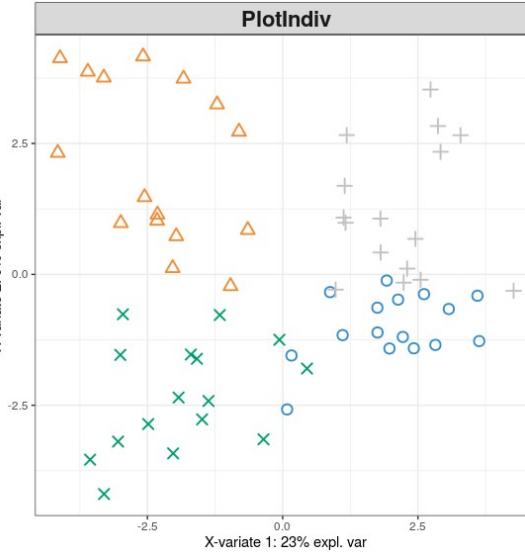
Contribution on comp 1



Contribution on comp 2

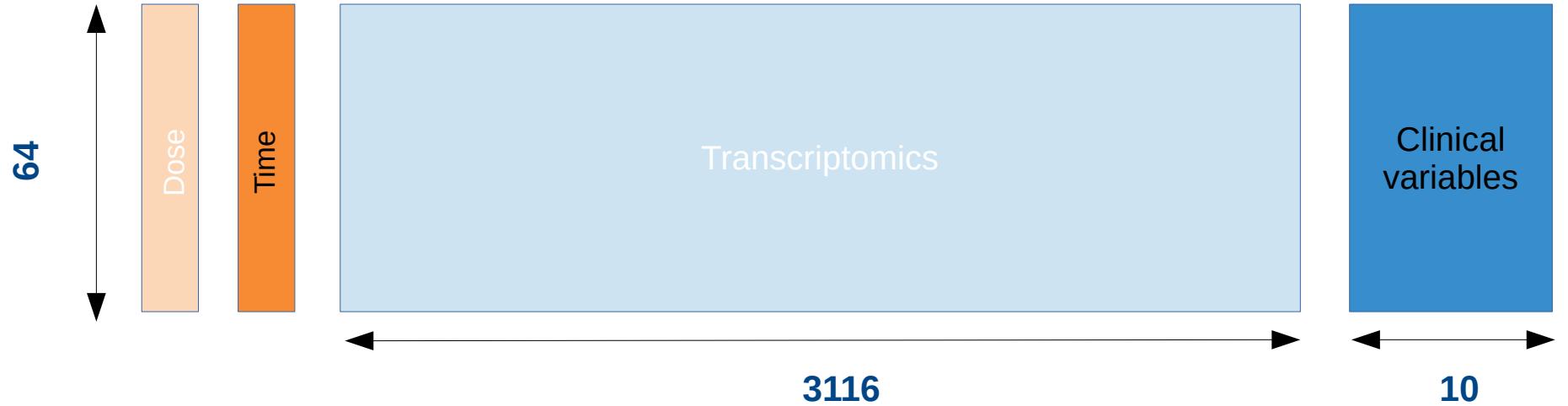


S-PLS-DA



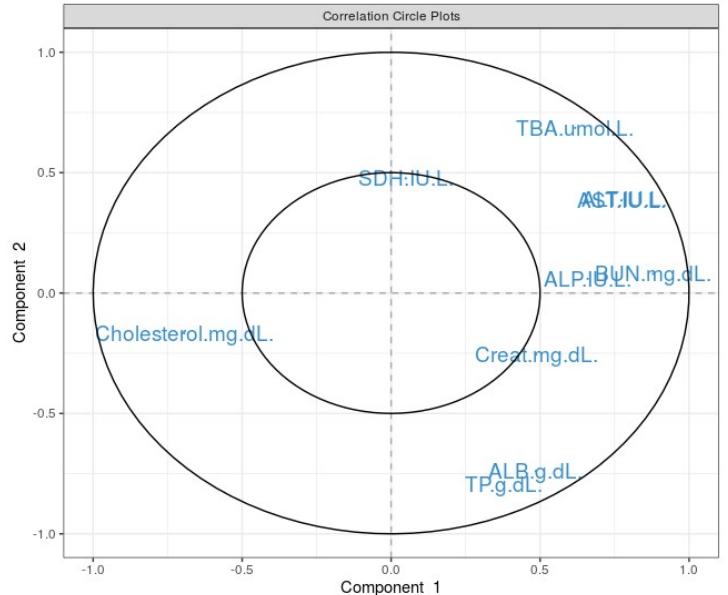
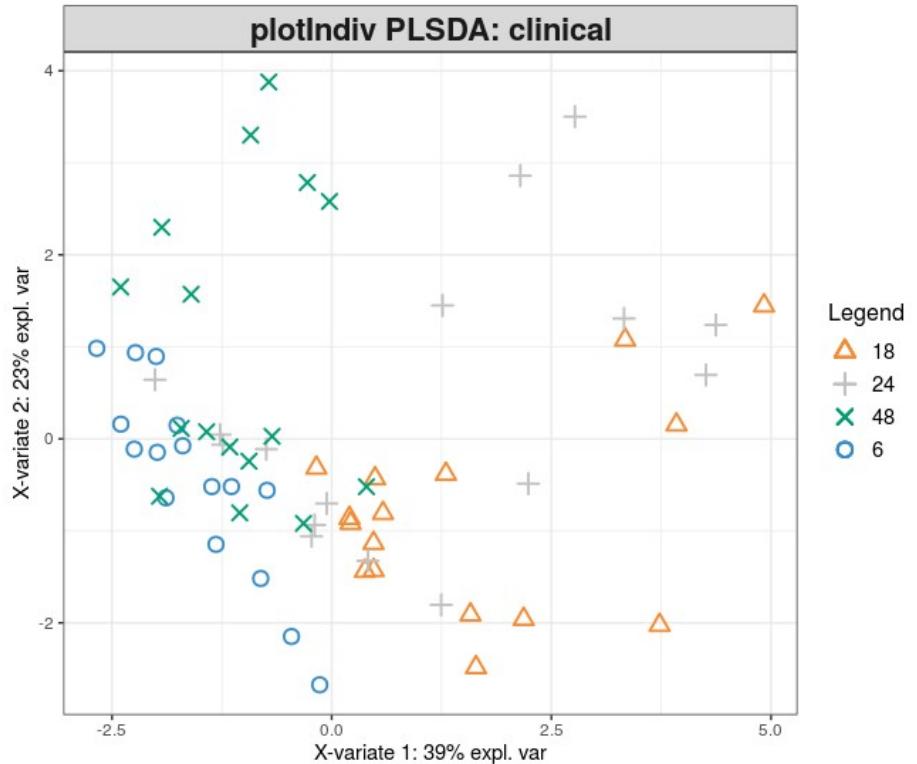
Answer: Probably, something to investigate...

Supervised analysis: clinical / time



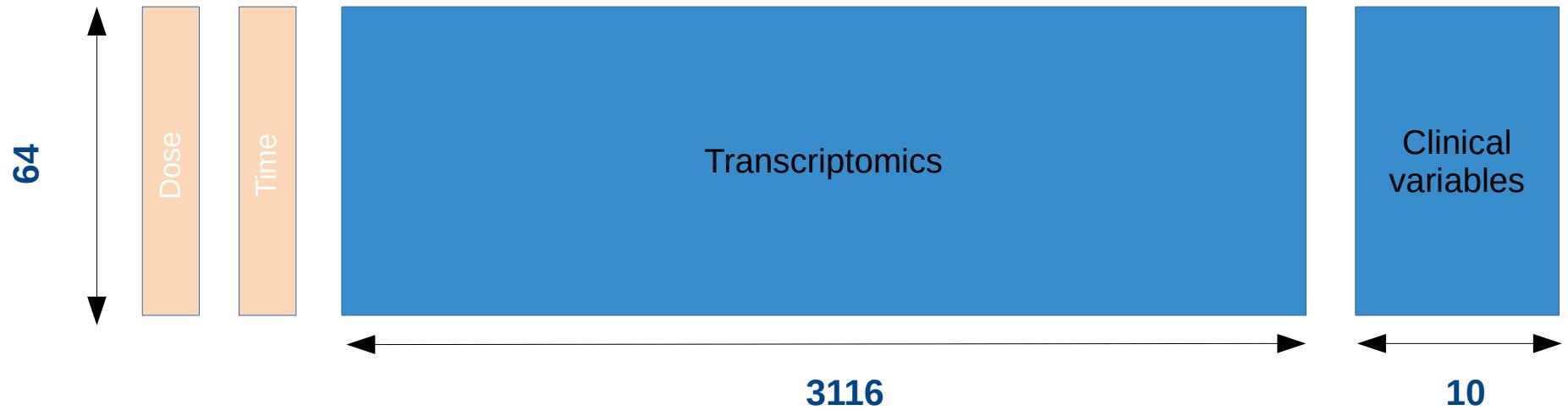
Question: Do we observe a better discrimination with the clinical data?

PLS-DA clinical / time



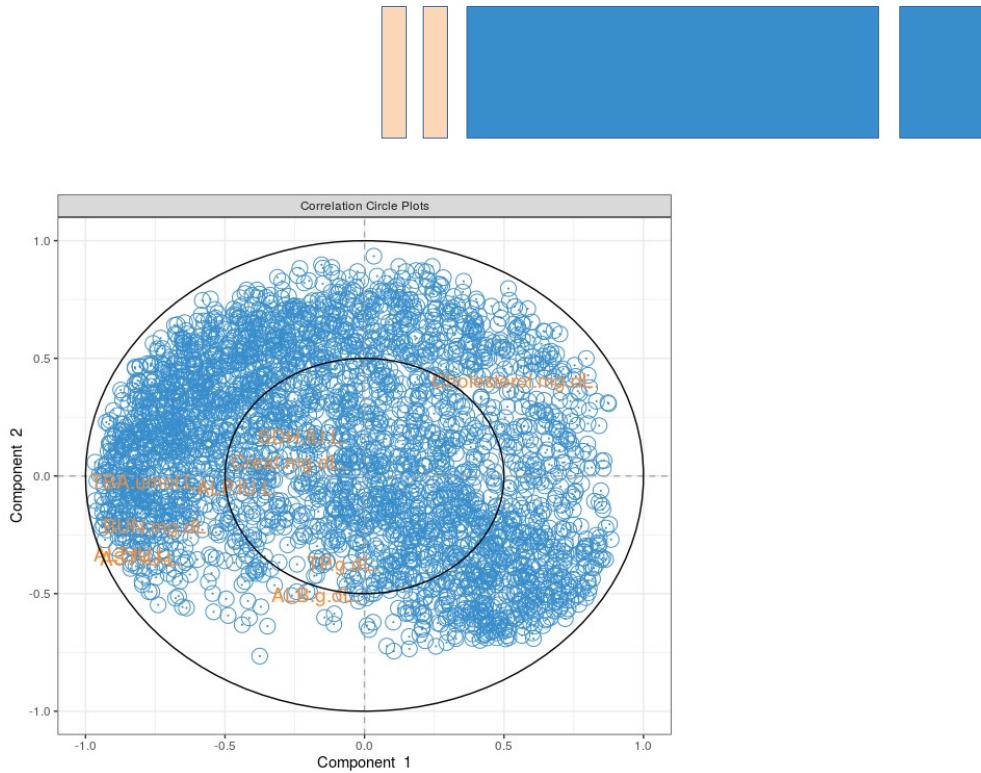
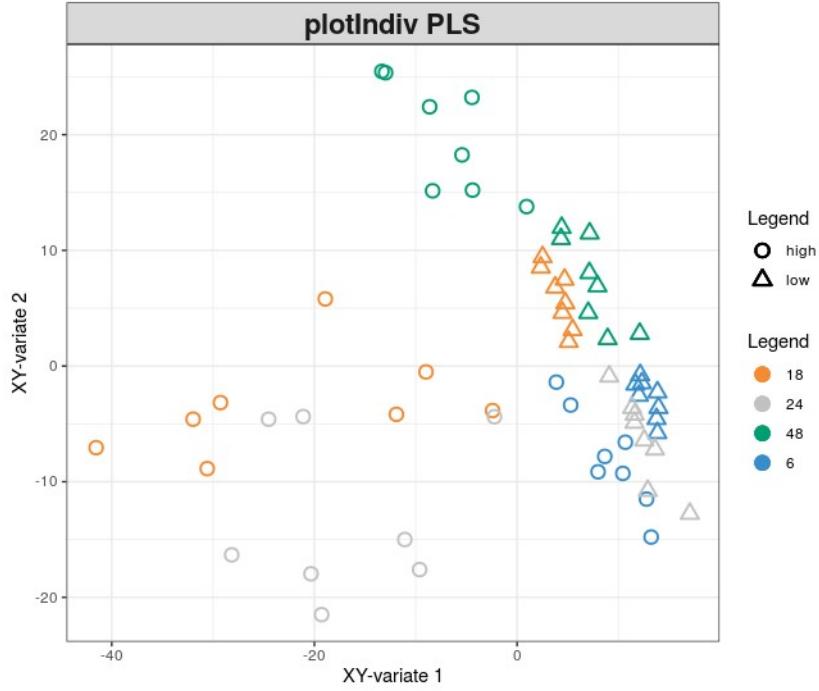
Answer: not as good as transcriptomics.

Unravel relationships between 2 datasets



Question: Can we unravel relationships between transcriptomics data and clinical data ?

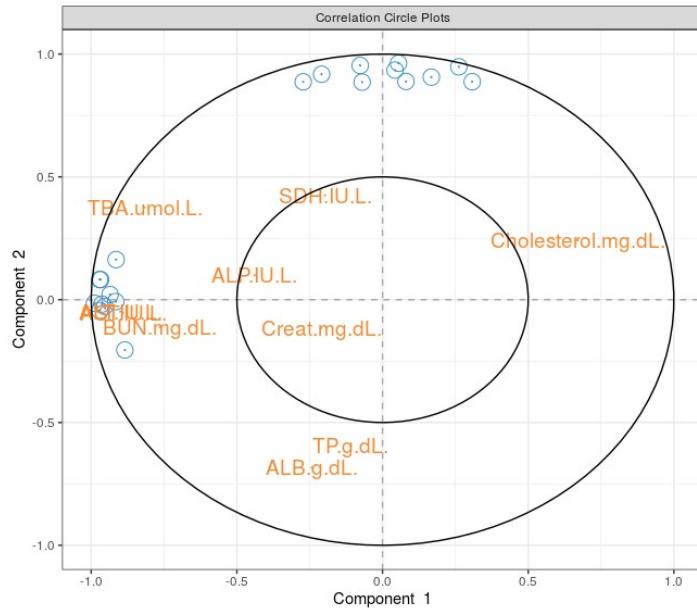
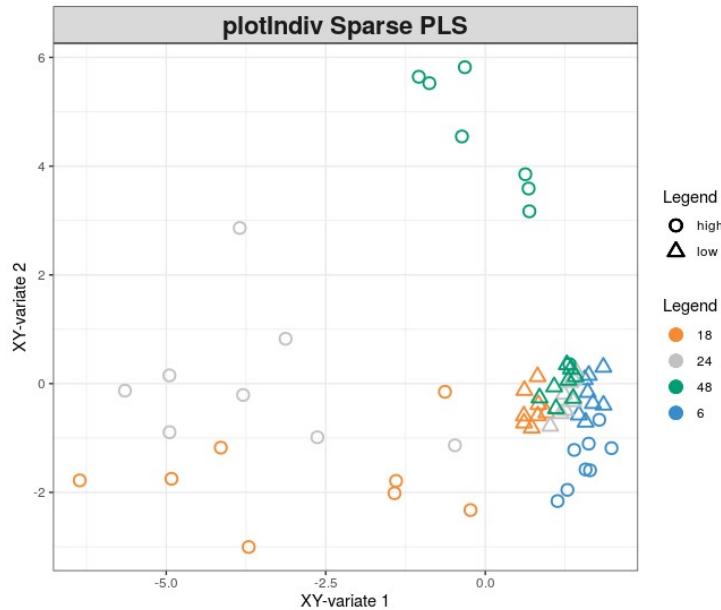
PLS: transcriptomics / clinic



Answer: well, well... to be investigated

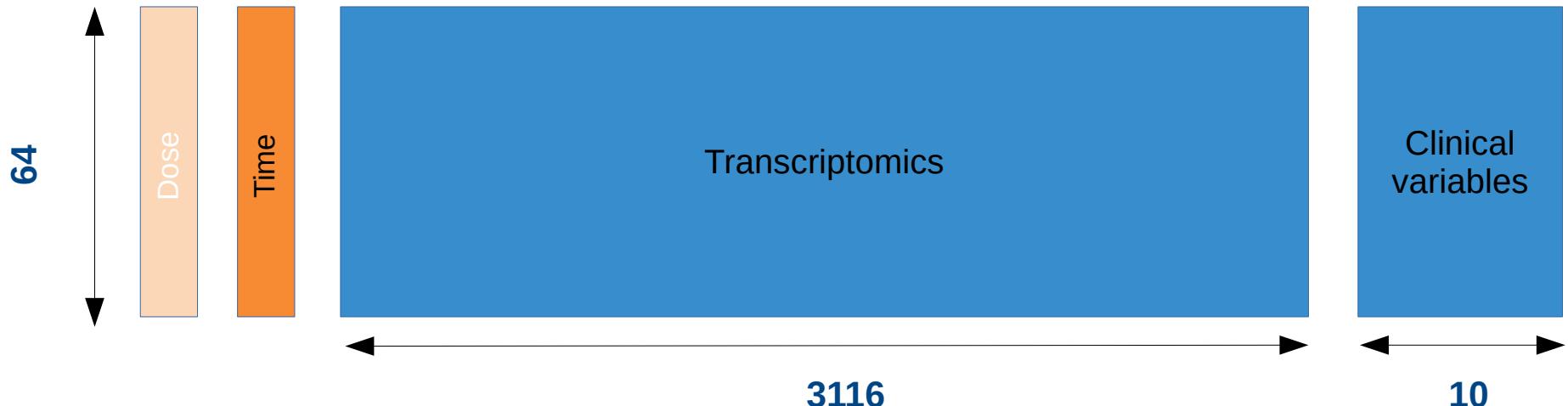
Sparse PLS: transcriptomics / clinic

Question: Can we unravel relationships between transcriptomics data and clinical data? **What are the genes that characterize these relationships?**



Answer: interesting trends on the individual plot and few genes involved.

Multi-blocks supervised analysis



Question: Does the integration of the clinical and transcriptomics datasets bring better insight into the biological similarities between samples within the same treatment dose?

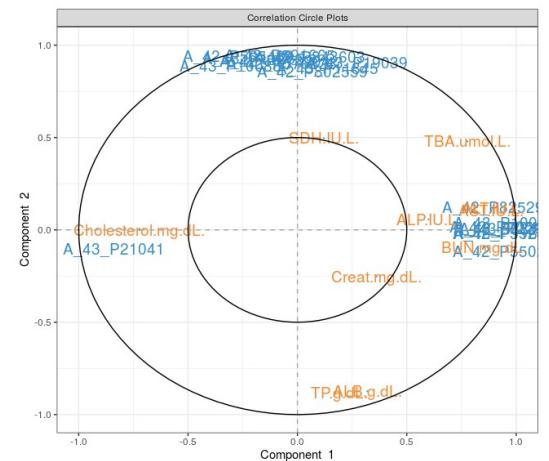
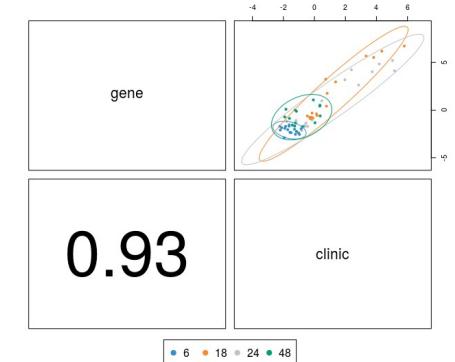
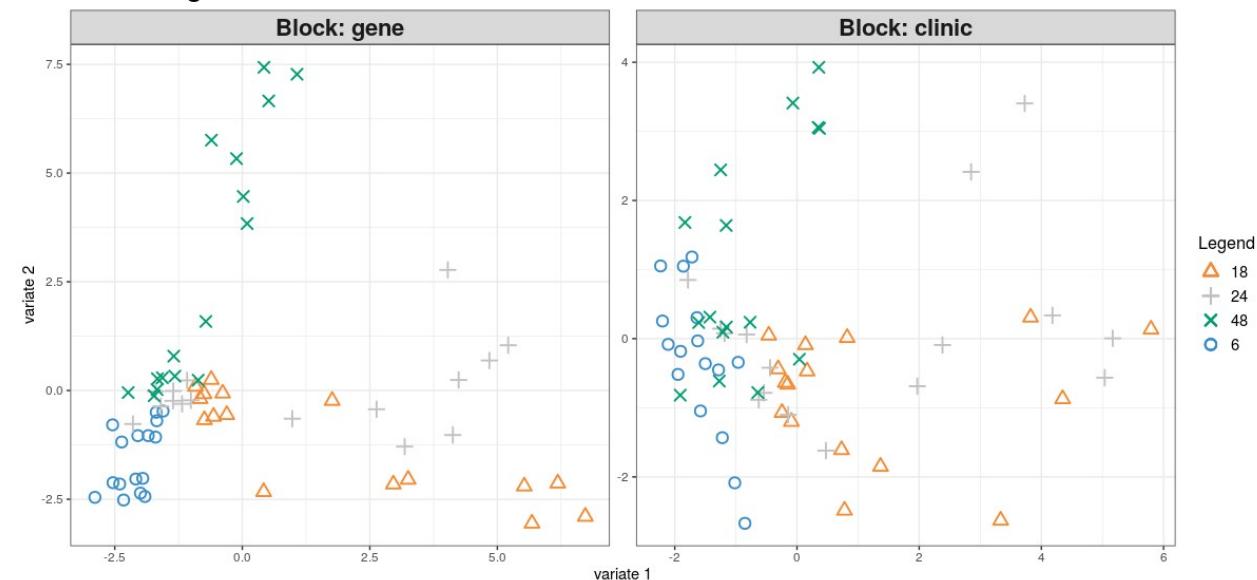
Investigation carried out
with two design matrices

	Full design			DA-oriented design			
	Tr.	Cl.	Time	Tr.	Cl.	Time	
Trans.	0	1	1	Trans.	0	0.1	1
Clinic.	1	0	1	Clinic.	0.1	0	1
Time	1	1	0	Time	1	1	0

Multi-blocks sparse PLS-DA: transcriptomics / clinic / time

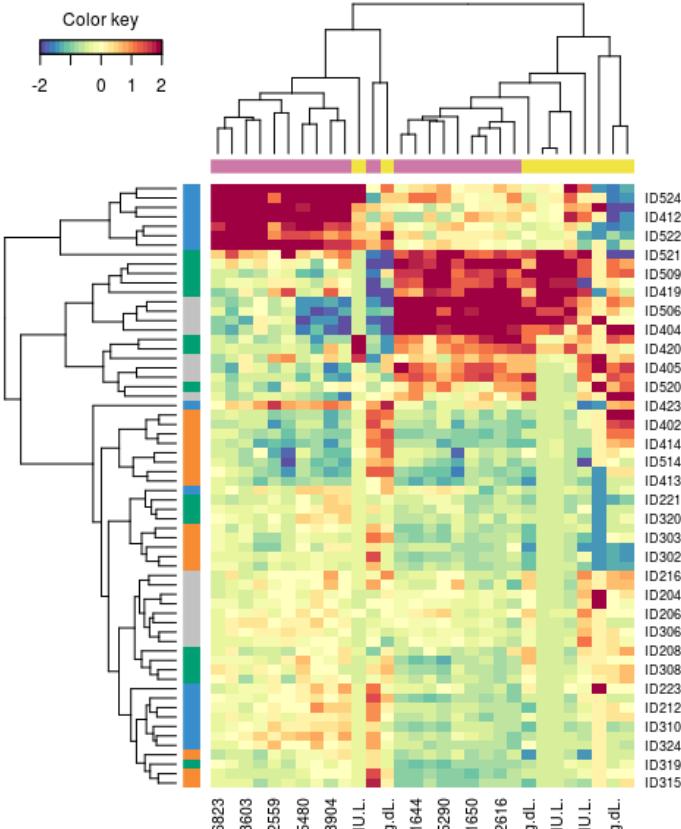


Full design

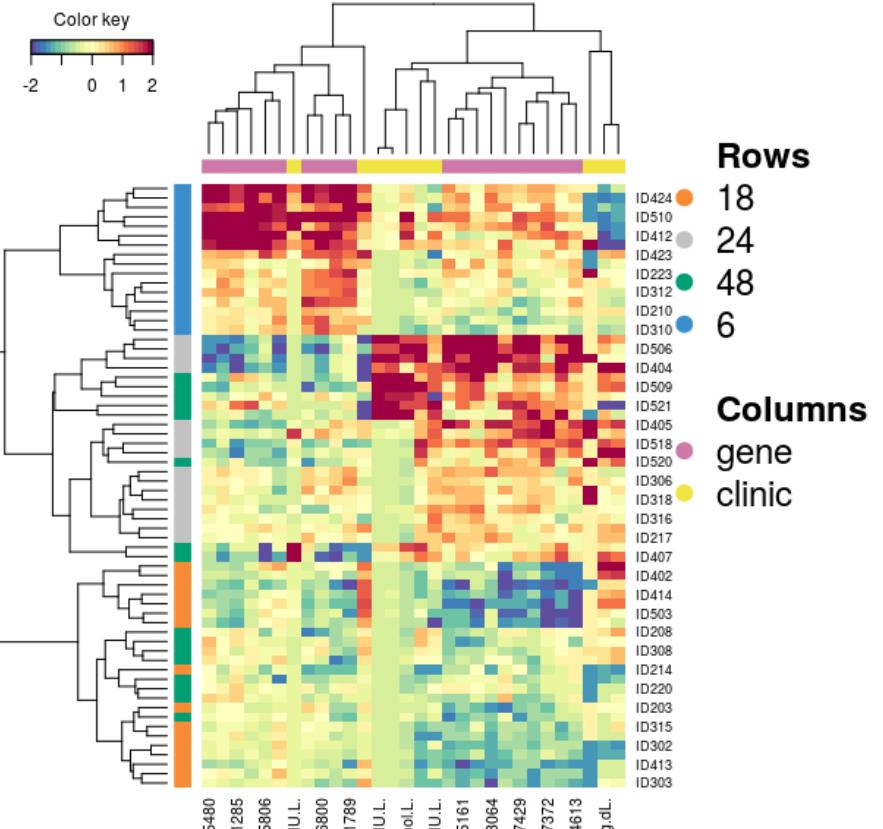


Answer: results to be investigated...

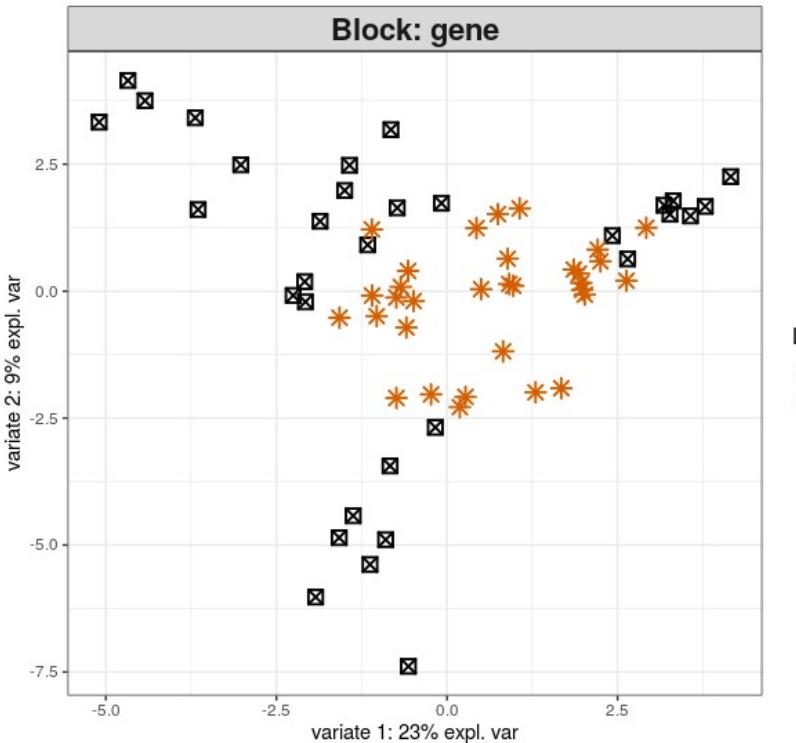
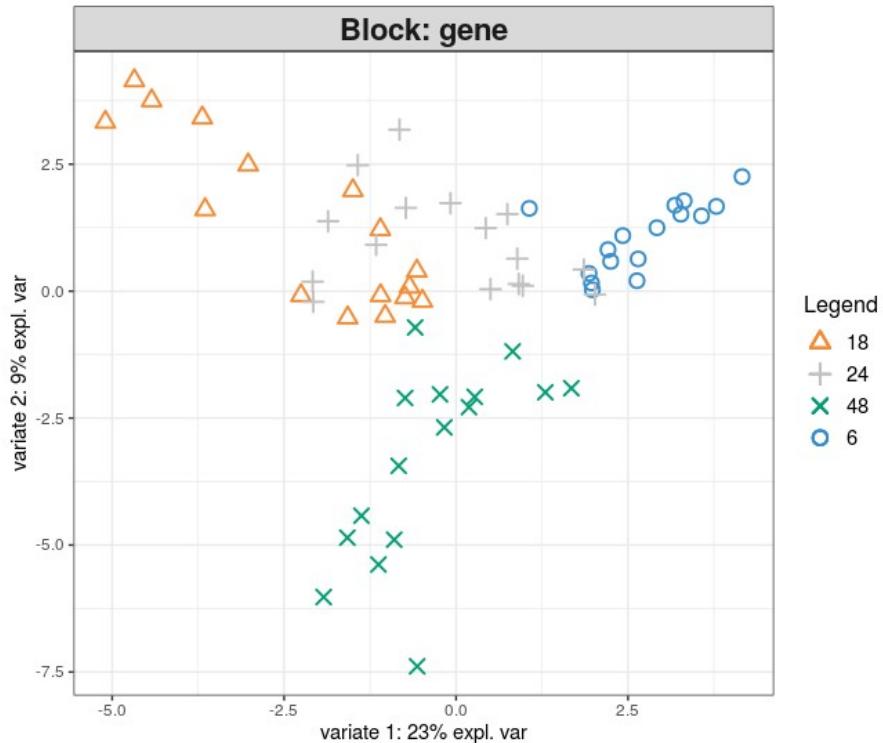
Multi-blocks sparse PLS-DA: transcriptomics / clinic / time



function
`cimDiabolo()`



Multi-blocks sparse PLS-DA: transcriptomics / clinic / time



DA-oriented design